

Constructional morphology of the shell/ligament system in opisthogyrate rostrate bivalves

J. Echevarría, S. E. Damborenea and M. O. Manceñido

CONICET – Museo de La Plata, Paseo del Bosque s/n, (1900) La Plata, Buenos Aires province, Argentina.

Email: javierechevarria@fcnym.unlp.edu.ar

ABSTRACT: The bivalve ligament provides the thrust for shell opening, acting as the resistance in a lever system against which adductor muscle effort is applied. Usually, its outer lamellar layer is subjected to tensile stress, while the inner fibrous layer is compressed, with the pivotal axis located between them. However, opisthogyrate rostrate bivalves display a concave dorsal margin, and both the umbo and the postero-dorsal angle of the shell project dorsally to the ligament, which then fails to act as pivotal axis. Three opisthogyrate rostrate genera of unrelated lineages show somewhat different solutions to this morpho-functional challenge. In *Cuspidaria* (Anomalodesmata), the ligament is internal, subjected only to compression and ventral to the pivotal axis, a thickened periostracum develops, forcing the dorsal margins of the valves to act as pivotal axis, and the posterior parts of the shell's dorsal margins gape dorsally. In *Nuculana* (Palaeotaxodonta), the inner layer of the ligament is internal, the outer layer is external but reduced, and some species develop a dorsal ridge parallel to the commissural plane, on a level with the rostrum and acting as pivotal axis. In *Pterotrigonia* (Palaeoheterodonta) and other rostrate trigoniids, the ligament is external opisthodontic, but is allometrically reduced. Trigoniids may also develop a dorsal ridge.

KEY WORDS: Allometry, Anomalodesmata, Bivalvia, concave dorsal margin, functional morphology, Nuculanida, Palaeoheterodonta, Palaeotaxodonta, Poromyida, Trigoniida.



In bivalve molluscs the ligament joins both valves dorsally and, being flexible, provides the thrust for opening the shell, acting as the resistance in a lever system against which contraction of adductor muscles acts as effort (Owen *et al.* 1953). The relative position of resistance, effort and fulcrum allows for the distinction of three classes of levers, only the first two being of interest here. In levers of class 1, the fulcrum is located between the two forces, which act in the same direction although at either side of the fulcrum (Fig. 1A). In levers of class 2, the resistance is located between the fulcrum and the effort, which acts in opposite direction to the resistance (Fig. 1B). Usually, in an external parivincular ligament, the outer lamellar layer is subjected to tensile stress, while the inner fibrous layer is exposed to compression, with the pivotal axis (representing the fulcrum of the lever system) located between them (Trueman 1949). The effort (i.e., the action of the adductor muscles) is always ventral to the pivotal axis; tensile stress develops dorsal to the pivotal axis and in the same direction as the effort, constituting the resistance of a class 1 lever, while compression develops ventral to the pivotal axis and in the opposite direction with respect to the effort, hence resulting in a class 2 lever (Fig. 1C). When a concave dorsal margin is developed, the pivotal axis is shifted dorsally to a line connecting the umbo and the elevated postero-dorsal angle of the shell, and so the tensile stress of the lamellar layer, now ventral to the pivotal axis, would prevent the valves from opening (Fig. 1D).

A prosogyrate shell shape, generally associated with the development of a lunule, is currently very frequent in shallow-burrowing bivalves, and is considered of adaptive value during burrowing (Stanley 1975a, b). On the other hand, an opisthogyrate shell shape is unusual amongst burrowing bivalves, with the main exception being the order Trigoniida (Palaeoheterodonta), where this condition is very common (Stanley 1977, 1978). The most extreme cases are the crescentic shells of many Pterotri-

giinae. These frequently have a produced posterior margin generating a rostrum, resulting in a concave escutcheon. Thus, the umbo and the postero-dorsal angle of the shell project dorsally relative to the ligament, which then fails to act as a pivotal axis.

A constructional morphology approach was employed in order to understand the mechanical operation of the shell–ligament system in the Pterotrigoniinae. Ligament structure and evolution have been extensively treated in the literature and their detailed discussion is beyond the scope of this contribution. *Neotrigonia* Cossmann, 1912 is the only extant trigoniids genus but, unfortunately, it strongly differs from *Pterotrigonia* v. Hoepen, 1929 in shell shape, as it is orthogyrate and triangular-shaped (Stanley 1978; Morton 1987a). Two other opisthogyrate rostrate extant genera of unrelated lineages were also analysed: *Nuculana* Link, 1807 (Palaeotaxodonta) and *Cuspidaria* Nardo, 1840 (Anomalodesmata). The rostrate, crescentic shape is infrequent in both Palaeotaxodonta and Anomalodesmata, suggesting a convergent development of this shell morphology in both lineages and in the trigoniids. These three taxa have few other common features: they differ in feeding habits, hinge and ctenidia, but they are all shallow burrowers. The particular shell morphology developed by all these taxa is probably related to this life habit: opisthogyrate growth favours an upward orientation of the posterior margin, making it easier for the animal to reach the water–sediment interface. The rostrum would thus allow the animal to reach the water column while most of the body remains below the sediment surface (Stanley 1977), in the same way as a snorkel allows a diver to maintain contact with the atmosphere while the body is underwater. Within the septibranchs, the rostrum is probably an adaptation for rapid eversion of the raptorial siphon (Morton 1981, 1987b). Also, the reduced size of the posterior end of the shell would reduce scour around it (Stanley 1977). Interestingly, both *Nuculana*

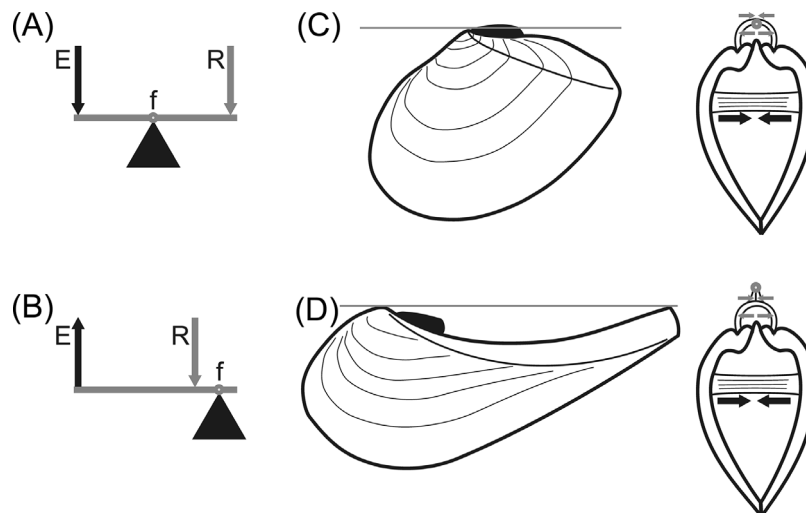


Figure 1 Bivalve shell as a lever system. (A–B) schematic drawings of class 1 (A) and class 2 (B) levers. Abbreviations: f = fulcrum; R = resistance; E = effort. (C–D) diagrams showing working of ligaments in prosogyrate (C) and opisthogyrate (D) shells. Grey line (on the left) and small grey circle (on the right) show the position of the pivotal axis; grey arrows indicate the direction of action of the resistance of both ligamental layers: lamellar (upper arrows, acting by tensile stress) and fibrous (lower arrows, acting by compression); black arrows indicate the direction of action of the effort, provided by the adductor muscles. Note in (D) that the resistance provided by the lamellar layer, being ventral to the pivotal axis (as is the effort) would prevent the valves from opening. Modified and extended from Yonge & Thompson (1976).

and *Cuspidaria* are siphonate species, unlike *Pterotrigonia*, and they both generate water currents by pumping action, due to muscular contractions of the ctenidia (Yonge 1928, 1939).

1. Material and methods

The following trigoniide samples were analysed:

Scabrotrigonia thoracica (Morton, 1834): USNM-PAL 636024, USNM-PAL 636032, USNM-PAL 129702, USNM-PAL 130844, USNM-PAL 386648.

Scabrotrigonia eufaulensis (Gabb, 1860): USNM-PAL 636026, USNM-PAL 27923, USNM-PAL 28435, USNM-PAL 305123.

Myophorella garatei Leanza, 1981: MCF-PIPH 427–429, MOZ-PI 930, MOZ-PI 933, MOZ-PI 7192–7194.

Myophorella coihuicoensis (Weaver, 1931): MCF-PIPH 484, MCF-PIPH 486, MCF-PIPH 488–489, MCF-PIPH 491–492, MOZ-PI 1687, MOZ-PI 7186–7188, MOZ-PI 8974–8975, MOZ-PI 8977–8978.

Myophorella volkheimeri Leanza & Garate, 1987: MCF-PIPH 459, MOZ-PI 1752, MOZ-PI 4173, MOZ-PI 7197–7198, MOZ-PI 7201–7204, MOZ-PI 8979.

Pterotrigonia coheni Leanza, 1993: MCF-PIPH 385, MCF-PIPH 389.

Pterotrigonia transatlantica (Behrendsen, 1892): MOZ-PIPH 2953, MOZ-PIPH 3072, MOZ-PIPH 3189, MOZ-PIPH 4397, MOZ-PIPH 4400.

Steinmanella caicayensis Lazo & Luci, 2013: MCF-PIPH 410, MCF-PIPH 412, MLP-PI 34629–34630.

Neotrigonia spp.: USNM 41485, USNM 88769, USNM 162080, USNM 465397, USNM 637181, USNM 76059, USNM 703373, USNM 162081, USNM 681695, USNM 846264, USNM 198491, USNM 75120, USNM 126852.

In each case, nymph length was measured with callipers and was compared to shell height (less affected by rostration than length); given the strong similarity in shell morphology

amongst extant species of *Neotrigonia*, they were pooled together for the purpose of this analysis. Both variables were log-transformed and a standardised major axis regression (SMA) was fitted to the dataset; the slope of that regression is the allometric coefficient. An allometric coefficient of 1 implies isometry, whilst greater or lower values imply positive allometry (i.e., relative ligament increase) or negative allometry (i.e., relative ligament decrease), respectively. For a discussion of line-fitting in the context of allometry, see Warton *et al.* (2006). Allometry analyses were performed using PAST 2.17 (Hammer *et al.* 2001).

Materials of the extant species *Nuculana inaequisculpta* (Lamy, 1906) (MLP-Ma 12389), *Nuculana sulculata* (Gould, 1852) (MLP-Ma 12386, MLP-Ma 12391, MLP-Ma 12399), *Nuculana* sp. (MLP-Ma 9120), *Cuspidaria tenella* Smith, 1907 (MLP-Ma 7382) and *Cuspidaria* sp. (MLP-Ma 7381) were analysed, paying special attention to the dorsal margin, the ligament and shell alignment during opening of the valves. Comparisons with examples from the literature were also made.

Institutional repositories. MCF-PIPH, Museo Paleontológico Carmen Funes, Plaza Huinacul, Argentina; MLP-Ma, Malacology, Invertebrate Zoology Collection, La Plata Natural Sciences Museum, Argentina; MLP-PI, Invertebrate Palaeontology Collection, La Plata Natural Sciences Museum, Argentina; MOZ-PI, Museo Provincial de Ciencias Naturales “Dr. Prof. Juan A. Olsacher”, Zapala, Argentina; USNM, United States National Museum of Natural History (NMNH), Department of Invertebrate Zoology, Smithsonian Institution, Washington DC, USA; USNM-PAL, Department of Paleobiology, Smithsonian Institution, Washington DC, USA.

2. Results

2.1. Shell/ligament system in *Nuculana*

Both layers in the ligament of *Nuculana* differ in position and relative development. The fibrous layer is strongly developed ventrally, placed in a resiliifer (Fig. 2A–B, res.) and acting as

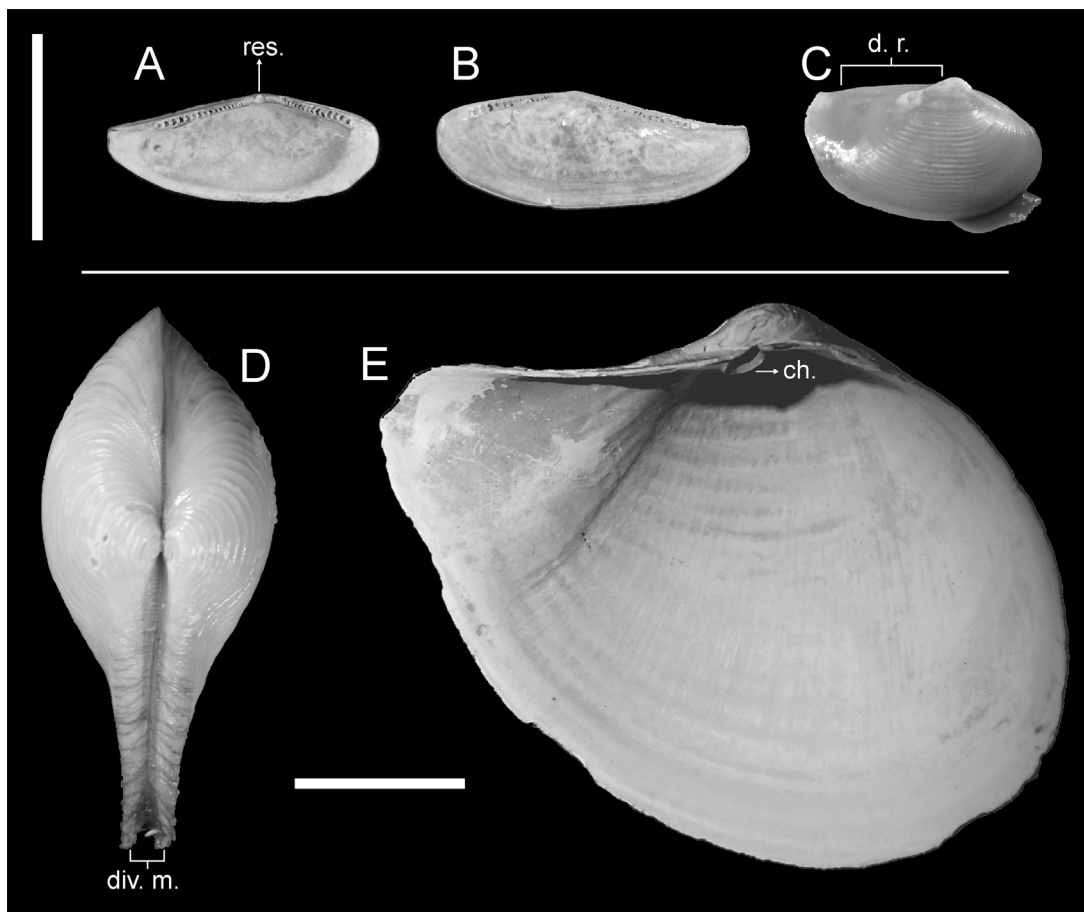


Figure 2 Part of the analysed material for extant species of nuculanids and cuspidariids. (A–B) MLP-Ma 9120, internal view of *Nuculana* sp.: (A) left valve; (B) right valve. (C) MLP-Ma 12389, right lateral view of *N. inaequisculpta*. (D) MLP-Ma 7381, dorsal view of *Cuspidaria* sp. (E) MLP-Ma 7382, internal view of left valve of *C. tenella*. Abbreviations: ch. = chondrophore; d. r. = dorsal ridge; div. m. = divergent margins; res. = resifier. Scale bars = 1cm.

an internal ligament, while the lamellar layer is still external but strongly reduced (Owen 1959) and, hence, does not disturb shell opening. Also, part of the lamellar layer develops anteriorly to the umbo, following the displacement of the pivotal axis from the dorsal margin; in fact, protobranchs (with the exception of the Solemyoidea) are the only bivalves in which the ligament grows anterior-ward (Waller 1990). All resistance results from compression only, ventral to the pivotal axis, so this is a class 2 lever.

Although the genus is usually characterised as having a rostrate posterior end, it shows a high interspecific variability of this character; whilst some species bear a short pointed rostrum (associated with an almost straight dorsal margin), others have a long and recurved one (Coan *et al.* 2000; Coan & Valentich-Scott 2012). In the latter case, the dorsal margin is concave, as in *N. pernula* (Müller, 1779), *N. navisa* (Dall, 1916) or *N. hamata* (Carpenter, 1864); sometimes the rostrum can be extremely long, as in *N. extenuata* (Dall, 1897) (Coan *et al.* 2000). Some species may also bear heavy, widely-spaced commarginal ribs (Coan *et al.* 2000), recalling somewhat the strong ornamentation of trigoniids; and *N. cordyla* (Dall, 1908) even has transverse ribs on the rostrum (Coan & Valentich-Scott 2012). Nevertheless, unlike trigoniids, most species of *Nuculana* are usually small, rarely exceeding 2 cm in length. *Nuculana inaequisculpta* also has a dorsal ridge parallel to the commissural plane, thus resulting in a straight dorsal margin levelled with, or even surpassing, the rostrum and acting as pivotal axis (Fig. 2C, d.r.).

Other genera amongst the Nuculanoidea may also develop a rostrum, though rarely associated with a concave dorsal margin; but *Yoldia cooperii* Gabb, 1865 and *Scaeolea illepada* Iredale, 1929 are exceptions (Abbott & Dance 1982; Coan *et al.* 2000). Many species of *Saccella* Woodring, 1925 may also bear a concave dorsal margin (Coan & Valentich-Scott 2012). Another interesting rostrate Nuculanoidea is the fossil genus *Ryderia* Wilton, 1830, which not only has a very long rostrum (with a concave dorsal margin), but also frequently shows divaricate ornamentation (Hodges 2000).

2.2. Shell/ligament system in *Cuspidaria*

Before describing the shell/ligament system, it should be noted that most cuspidariids are relatively small bivalves (<5 mm long); sizes at which shells can be relatively flexible, so that valve opening would not pose major problems. Something similar occurs in *Nuculana*, although in this genus shells are usually more heavily calcified. Since Pterotrigninae are generally medium-sized bivalves, large shells of *Cuspidaria* were preferred for the analysis.

Ligament in *Cuspidaria* is completely internal, with both lamellar and fibrous layers settled in a chondrophore (Fig. 2E, ch.); a lithodesma placed ventral to the ligament avoids expansion of the ligament during valve closure (Yonge 1978; Yonge & Morton 1980). Thus, the ligament can act only by compression, both layers acting ventrally to the pivotal axis and in an opposite direction to the effort, resulting effectively in a class 2 lever. But there is another peculiarity in *Cuspidaria*, which is

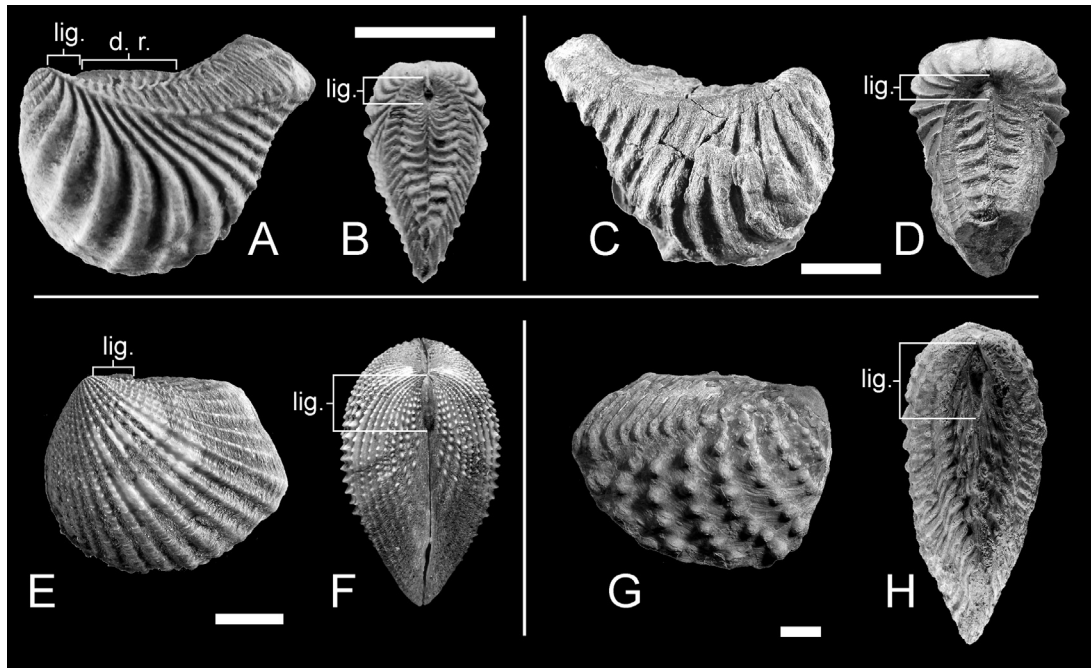


Figure 3 Part of the analysed material of trigonioids. (A–B) *Myophorella garatei*, from the Valanginian–lower Hauterivian of Argentina: (A) MCF-PIPH 428, left lateral view; (B) MCF-PIPH 427, dorsal view. (C–D) *Pterotrigonia coheni*, from the Tithonian–Valanginian of Argentina: (C) MCF-PIPH 389, right lateral view; (D) MCF-PIPH 385, dorsal view. (E–F) USNM 465397, *Neotrigonia margaritacea* (Lamarck, 1804), from Australia: (E) left lateral view; (F) dorsal view. (G–H) MCF-PIPH 410, *Steinmanella caicayensis*, from the Valanginian of Argentina: (G) right lateral view; (H) dorsal view. Abbreviations: d. r. = dorsal ridge; lig. = ligament/nymph. Scale bars = 1 cm.

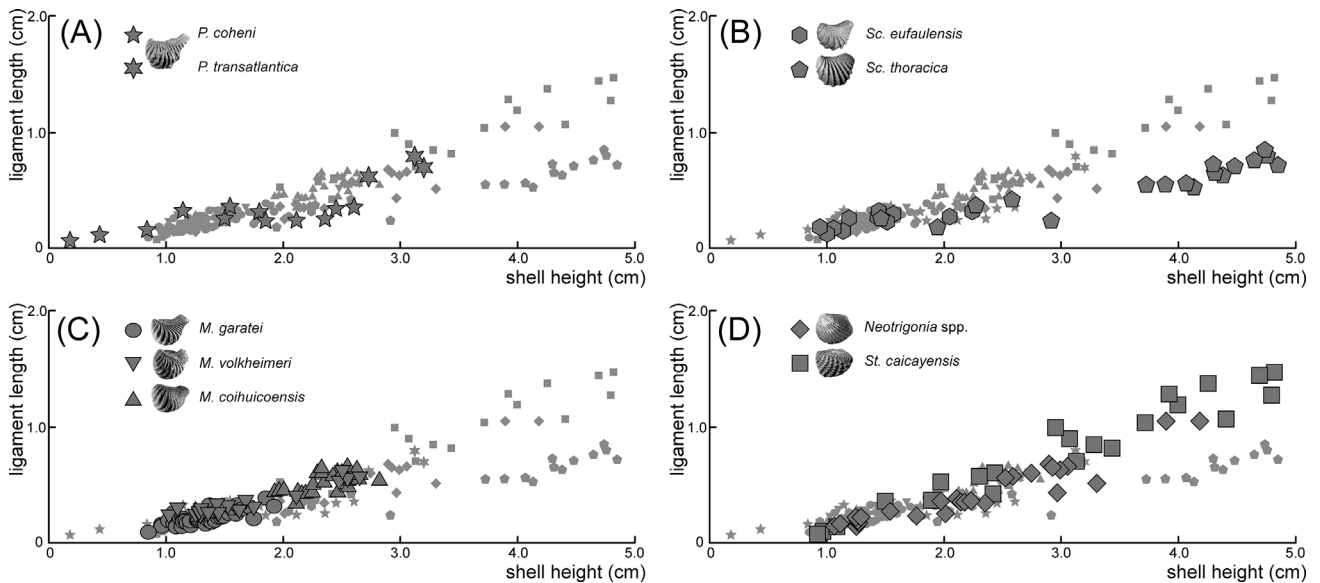


Figure 4 Dispersion plots showing the length of ligament (*y*-axis) against height of shell (*x*-axis) for the analysed species of trigonioids: (A) *Pterotrigonia* species; (B) *Scabrotrigonia* species; (C) *Myophorella* species; (D) orthogyrate species. Plots of the other species are shown in all graphs as small grey symbols for easier comparison.

the development of a thickened periostracum extending across the entire dorsal margin of the shell (Yonge 1978; Yonge & Morton 1980). This structure results from the fusion of the periostracum of both valves and, as a consequence of its development, the dorsal margin has to act as pivotal axis. To avoid obstruction during shell aperture, dorsal margins diverge and gape from the sagittal plane on the portion of the rostrum which projects dorsally (Fig. 2D, div. m.). This feature also allows for a more efficient aperture of the valves, since the

closeness of the pivotal axis to the resistance (i.e., the ligament) renders maximum advantage of its thrust.

Just as in nuculanids, the rostrum within septibranchs is not restricted to the genus *Cuspidaria*. *Acreuciroa* Thiele in Thiele & Jaekel, 1931, *Austroneaera* Powell, 1937, *Luzonia* Dall & Smith, in Dall 1889, *Myonera* Dall & Smith, in Dall 1886, *Plectodon* Carpenter, 1864, *Bathyneaea* Scarlato & Starobogatov, 1983 and *Cardiomya* Adams, 1864 also have a produced posterior margin, although only in the last three it is

Table 1 Results of the allometry analyses of ligament length (y -variable) vs. shell height (x -variable) for the trigoniide species considered. Asterisks indicate values significantly departing from isometry; values lower than one indicate negative allometric trend (i.e., relative ligament length reduction). Abbreviations: al.coef. = allometric coefficient; $p_{(al. coef.=1)}$ = probability of the allometric coefficient being one; N = number of specimens.

species	al. coef.	$p_{(al. coef.=1)}$		N
<i>P. coheni</i>	0.62	0.001	*	12
<i>P. transatlantica</i>	–	–		4
<i>M. garatei</i>	1.59	0.001	*	42
<i>M. volkheimeri</i>	1.10	0.325		19
<i>M. coihuicoensis</i>	1.21	0.032	*	29
<i>Sc. thoracica</i>	1.28	0.084		17
<i>Sc. eufalensis</i>	1.11	0.355		14
<i>Neotrigonia</i> spp.	1.26	0.002	*	32
<i>St. caicayensis</i>	1.50	<0.001	*	29

accompanied by a concave dorsal margin (Coan *et al.* 2000). Within the Cuspidarioidea, it is noteworthy that the shells of members of the family Spheniopsidae Gardner, 1928, slightly prosogyrate, lack a rostrum or at most, are modestly produced (Coan & Valentich-Scott 2012; Machado & Passos 2015; Morton *et al.* 2016).

2.3. Shell/ligament system in trigoniides

As an important difference from previously described groups, ligament in trigoniides is always external and parivincular (Newell & Boyd 1975; see also Fig. 3), hence it cannot act as a class 2 lever, because its lamellar layer will be always subjected to tensile stress during valve closure.

A dorsal ridge, such as the one present in *Nuculana inaequisculpta*, is commonly present in rostrate trigoniides (Fig. 3A, d.r.). Nevertheless, it is not as well developed as in the protobranch species and, in most cases, the rostrum projects dorsally beyond the ridge, still hindering shell opening. Shell edges, unlike those in *Cuspidaria*, remain parallel and in contact throughout the whole dorsal margin (Fig. 3B), so the straight margin of the dorsal ridge cannot act as pivotal axis.

When compared to shell height, the ligament seems to be shorter in rostrate opisthogyrate species (Fig. 4A–C) than in orthogyrate ones (Fig. 4D). Ontogenetic development shows a negative allometric relationship for these characters in *Pterotrigonia coheni* and a positive one for *M. garatei*, *M. coihuicoensis*, *Steinmanella caicayensis* and *Neotrigonia* spp., and possibly for *S. thoracica* as well (Table 1).

3. Discussion

3.1. Internal ligament as a solution to the shell opening problem

In rostrate opisthogyrate bivalves, the pivotal axis of the shell is shifted dorsally. If the lamellar layer of the ligament were to remain ventral to the pivotal axis, tensile stress would prevent shell opening (Fig. 1D). The most efficient solution to this problem would be the development of an internal ligament. In this way, the resilifer would press on the ligament, which would then act only by compression, the whole system acting effectively as a class 2 lever.

Nevertheless, the presence of an internal ligament in *Nuculana* and *Cuspidaria* should not be considered as an adaptation resulting from their particular morphology, but most likely as an exaptation (*sensu* Gould & Vrba 1982). Non-rostrate Nuculanidae and Cuspidariidae also bear internal ligaments, suggesting that this condition developed earlier, allowing for the acquisition of a crescentic shell shape.

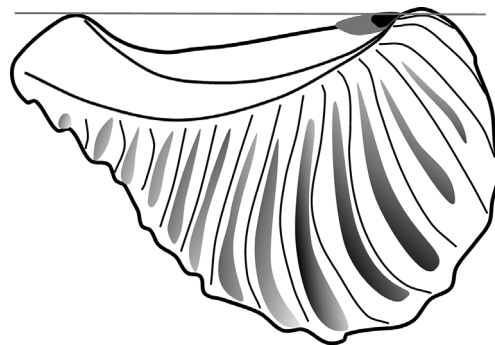


Figure 5 Sketch of *Pterotrigonia* showing the relationship between the pivotal axis of the shell (grey line) and reconstructed ligaments of different lengths (short ligament in black against long ligament in grey).

3.2. Shell opening in rostrate trigoniides

Despite the general variability in ligament length and development (Fig. 4; Table 1), it is clear that more rostrate trigoniides have shorter ligaments than less rostrate ones. This is true whether comparing genera (i.e., *Pterotrigonia* (Fig. 4A) and *Scabrotrigonia* (Fig. 4B) vs. *Steinmanella* and *Neotrigonia* (Fig. 4D)) or species within a genus (i.e., *M. garatei* vs. *M. volkheimeri* and *M. coihuicoensis* (Fig. 4C)). Also, it is noteworthy that species having longer rostra tend to show isometric or even negative allometric relationships to size (with the main exception of *M. garatei*).

On functional grounds, this could have several meanings. Stanley (1977, 1978) suggested that in *Neotrigonia*, the muscular foot aids the ligament during shell opening, due to the large amount of friction in the hinge system. It is possible, then, that in rostrate trigoniides, the foot was mainly responsible for shell opening, whilst ligament reduction would reduce the disturbance of tensile stress. On the other hand, the ligament is not only reduced, but is also very close to the umbo (though always opisthodic; Fig. 3). Although the antero-posterior axis of the ligament does not coincide with the pivotal axis of the shell, they are closer to each other on the anteriormost part of the dorsal margin, thanks to the development of the dorsal ridge. As a consequence, part of the ligament would develop dorsally to the pivotal axis, especially its anterior part (Fig. 5); this will be of particular importance if we consider the observation by Trueman (1949) that the anterior part of the ligament is the most effective for shell opening. As the ligament grows longer (posteriorly), most of it would remain ventral to the pivotal axis. So a short ligament would ensure that the functional part of the lamellar layer is raised dorsal to the pivotal axis, still assisting in shell opening.

3.3. Constructional aspects of the ligament in opisthogyrate shells

The relative shortening of the ligament in rostrate opisthogyrate trigoniides can also be explained on morphogenetic grounds. In orthogyrate bivalves, the direction of growth at any region of the mantle edge can be resolved into a radial component, radiating from the umbo and within the commissural plane (Fig. 6A, R), and a transverse component, normal to the commissural plane (Fig. 6B, N); both of them will be greatest mid-ventrally and least mid-dorsally (Wilbur & Owen 1964). In prosogyrate and opisthogyrate shells, a third component is added (Fig. 6C, T), acting tangentially to, and in the plane of, the commissure (Wilbur & Owen 1964). In opisthogyrate shells, tangential and radial components will augment one another anterior to the umbos, with a larger resultant growth; whilst posterior to them, they will oppose (Fig. 6C). As a consequence, an allometric reduction of the dorsal margin

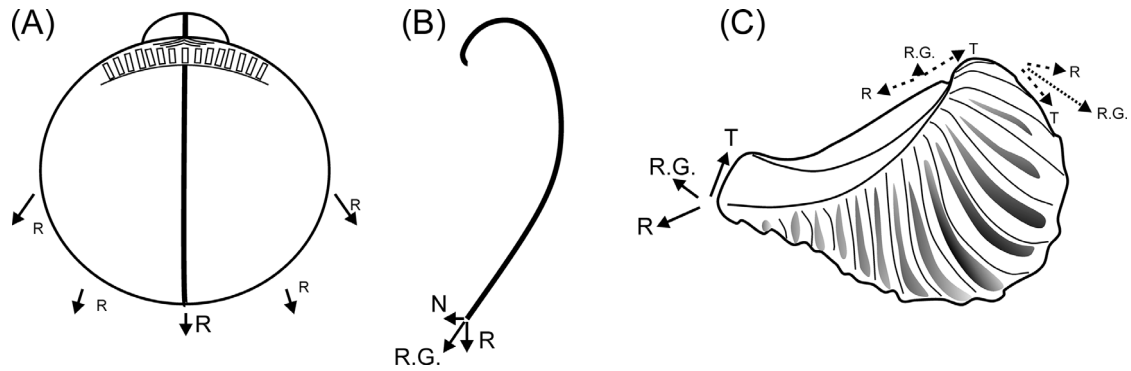


Figure 6 Diagrams showing growth components of bivalve shells. (A–B) growth on orthogyrate shells (modified from Wilbur & Owen 1964): (A) lateral view; (B) transverse section of one valve. (C) growth on an opisthogyrate shell exemplified on a right valve of *Pterotrignonia*. Abbreviations: N = transverse component, R = radial component, R.G. = resultant growth, T = tangential component.

(and of the ligament associated with it) would be expected, as opisthogyrate growth becomes stronger.

Thus, the reduction of the ligament in rostrate, crescentic shells can be interpreted as a constructional consequence of opisthogyrate shell growth and, given its functional value as discussed above, it should also be considered as an exaptation.

4. Conclusions

- In rostrate opisthogyrate bivalves, the umbo and the postero-dorsal angle of the shell project dorsally relative to the ligament, which then fails to act as a pivotal axis. This challenge, present in several bivalve groups, has been functionally solved in different, and sometimes complementary, ways.
- The presence of an internal ligament, acting by compression, results in a class 2 lever, which allows for the development of a rostrate shell morphology with a concave dorsal margin (exaptation in Nuculanidae and Cuspidariidae). The development of a dorsal ridge parallel to the commissural plane (in Nuculanidae), and the development of divergent margins on the posterior part of the dorsal margin (in Cuspidariidae), also allow for shell opening in opisthogyrate rostrate bivalves.
- When the ligament is external, shell growth promotes the allometric reduction of the ligament; this fact, together with the development of a dorsal ridge, allowed the Pterotrignoniinae and other rostrate trigoniinids to maintain the functionality of the ligament, although shell opening was most likely aided by a strong muscular foot.

5. Acknowledgements

The authors wish to express their gratitude to the organisers of the Symposium ‘Form, Function and Palaeobiology’, S. F. Vizcaino, E. N. K. Clarkson and B. E. Schoenemann, for their invitation to participate. We also thank the staff of the visited institutions for their assistance: Departments of Invertebrate Zoology and of Paleobiology, NMNH, Washington DC; Servicio Nacional de Geología y Minería de Chile (SERNAGEOMIN), Santiago, Chile; Museo Olsacher, Zapala, Argentina; Museo Carmen Funes, Plaza Huincul, Argentina; División Zoología de Invertebrados, Museo de La Plata, Buenos Aires, Argentina. Comments from two anonymous reviewers helped to improve the manuscript.

6. References

- Abbot, R. T. & Dance, S. P. 1982. *Compendium of seashells*. New York: E. P. Dutton Inc. 411 pp.
- Adams, A. 1864. On the Species of *Neaera* found in the Seas of Japan. *Annals and Magazine of Natural History*, ser. 3 **13**, 206–09.
- Behrensden, O. 1892. Zur Geologie des Ostabhanges der Argentinischen Kordillere. *Zeitschrift der Deutsche Geologischen Gesellschaft* **43**, 1–42.
- Carpenter, P. P. 1864. Supplementary report on the present state of our knowledge with regard to the Mollusca of the west coast of North America. *British Association for the Advancement of Science, Report* **33**, 517–686.
- Coan, E. V., Valentich-Scott, P. & Bernard, F. R. 2000. Bivalve seashells of Western North America. *Santa Barbara Museum of Natural History Monographs* **2**, 764 pp.
- Coan, E. V. & Valentich-Scott, P. 2012. Bivalve seashells of tropical West America, parts 1 and 2. *Santa Barbara Museum of Natural History Monographs* **6**, 1258 pp.
- Cossmann, M. 1912. Sur l'évolution des Trigonies. *Annales de paléontologie* **7**, 59–84.
- Dall, W. H. 1886. Reports on the results of dredging, under the supervision of Alexander Agassiz, in the Gulf of Mexico (1877–78) and in the Caribbean Sea (1879–80), by the U.S. Coast Survey Steamer *Blake*, Lieutenant Commander C. D. Sigsbee, U.S.N., and Commander J. R. Bartlett, U.S.N., Commanding. XXIX. Report on the Mollusca. Part 1. Brachiopoda and Pelecypoda. *Harvard University, Museum of Comparative Zoology, Bulletin* **12**(6) 171–318.
- Dall, W. H. 1889. Scientific results of explorations US Fish Commission Steamer *Albatross*, VII. Preliminary report on the collection of Mollusca and Brachiopoda obtained in 1887–88. *US National Museum, Proceedings* **12**, 217–362.
- Dall, W. H. 1897. Notice on some new or interesting species of shells from British Columbia and the adjacent region. *Natural History Society of British Columbia, Bulletin* **2**, 1–18.
- Dall, W. H. 1908. Reports on the dredging operations off the west coast of Central America to the Galapagos, to the west coast of Mexico, and in the Gulf of California, in charge of Alexander Agassiz, carried out by the U.S. Fish Commission Steamer *Albatross* during 1891, Lieutenant Commander Z. L. Tanner, commanding. XXXVII. *Bulletin of the Museum of Comparative Zoology at Harvard College* **43**, 205–487.
- Dall, W. H. 1916. *Checklist of the Recent bivalve mollusks (Pelecypoda) of the northwest coast of America from the Polar Sea to San Diego, California*. Los Angeles (Southwest Museum). 44 pp.
- Gabb, W. M. 1860. Description of new species of American Tertiary and Cretaceous fossils. *Journal of the Academy of Natural Science of Philadelphia* **4**(4), 375–406.
- Gabb, W. M. 1865. Description of new species of marine shells from the coast of California. *California Academy of Sciences, Proceedings* **3**, 182–90.
- Gardner, J. 1928. The molluscan fauna of the Alum Bluff Group of Florida. 5. Tellinacea, Solenacea, Mactracea, Myacea, Molluscoidea. *US Geological Survey Professional Paper* **142**, 185–249.

- Gould, A. A. 1852. Mollusca and shells. US Exploring Expedition during the years 1838, 1839, 1840, 1841, 1842 under the command of Charles Wilkes. *U.S.N. Reports* **12**, 1–498.
- Gould, S. J. & Vrba, E. S. 1982. Exaptation- A missing term in the science of form. *Paleobiology* **8**, 4–15.
- Hammer, Ø., Harper, D. A. T. & Ryan, P. D. 2001. PAST: Paleontological Statistics software package for education and data analysis. *Palaeontologia Electronica* **4**, 9 pp.
- Hodges, P. 2000. The Early Jurassic Bivalvia from the Hettangian and Lower Sinemurian of Southwest Britain. *Monograph of the Palaeontographical Society* **614**, 1–64.
- Hoepen, E. C. N. van, 1929. Die krytfauna van Soeloeland. 1. Trigonidae. *Paleontologiese Navorsing van die Nasionale Museum, Bloemfontein* **1**, 1–38.
- Iredale, T. 1929. Mollusca from the continental shelf of eastern Australia, Part 2. *Australian Museum Records* **17**(4) 157–89.
- Lamarck, J. P. B. 1804. Sur une nouvelle espèce de Trigonie, et sur une nouvelle d'Huitre découvertes dans le voyage du capitaine Baudin. *Annales du Muséum d'Histoire Naturelle* **4**, 353–54.
- Lamy, E. 1906. Lamellibranches recueillis par l'Expédition Antarctique Française du Dr. Charcot. *Bulletin du Muséum National d'Histoire Naturelle* **12**, 44–52.
- Lazo, D. G. & Luci, L. 2013. Revision of Valanginian Steinmanellinae bivalves from the Neuquén basin, west-central Argentina, and their biostratigraphic implications. *Cretaceous Research* **45**, 60–75.
- Leanza, H. A. 1981. Una nueva especie de *Myophorella* (Trigonidae-Bivalvia) del Cretácico Inferior de Neuquén, Argentina. *Ameghiniana* **18**(1–2), 1–9.
- Leanza, H. A. 1993. Jurassic and Cretaceous trigoniid bivalves from west-central Argentina. *Bulletins of American Paleontology* **105**(343), 1–95.
- Leanza, H. A. & Garate Zubillaga, J. I. 1987. Fauna de Trigonias (Bivalvia) del Jurásico y Cretácico Inferior de la provincia del Neuquén, Argentina, conservadas en el Museo Juan Olsacher de Zapala. In Volkheimer, W. (ed.) *Bioestratigrafía de los Sistemas Regionales del Jurásico y Cretácico de América del Sur* **1**, 201–55.
- Link, H. F. 1807. Beschreibung der Naturalien-Sammlung der Universität zu Rostock. *Abtheilung* **3**, 101–65.
- Machado, F. M. & Passos, F. D. 2015. Spheniopsidae Gardner, 1928 (Bivalvia): conchological characters of two new species from off Brazil, Southwestern Atlantic. *American Malacological Bulletin* **33**, 212–20.
- Morton, B. 1981. The Anomalodesmata. *Malacologia* **21**(1–2), 35–60.
- Morton, B. 1987a. The functional morphology of *Neotrigonia margaritacea* (Bivalvia: Trigonacea), with a discussion of phylogenetic affinities. *Records of the Australian Museum* **39**, 339–54.
- Morton, B. 1987b. Siphon structure and prey capture as a guide to affinities in the abyssal septibranch Anomalodesmata (Bivalvia). *Sarsia* **72**(1), 49–69.
- Morton, B., Machado, F. M. & Passos, F. D. 2016. The smallest carnivorous bivalve? Biology, morphology and behaviour of *Grippina coronata* (Anomalodesmata: Cuspidarioidea: Spheniopsidae) preying on epipsammic microcrustaceans in the southwestern Atlantic off Brazil. *Journal of Molluscan Studies* **82**, 244–58.
- Morton, S. G. 1834. *Synopsis of the organic remains of the Cretaceous Group of the United States*. Philadelphia: Key & Biddle. 88 pp.
- Müller, O. F. 1779. Zwoen wenig bekannten Muscheln, der Schinkenarhe und der gerunzelten Mahlermuschel. *Gesellschaft Naturforschender Freunde zu Berlin, Beschäftigungen* **4**, 55–59.
- Nardo, G. 1840. Sopra un nuovo genere di Conchiglie del mare Adriatico denominato *Cuspidaria*. *Atti Riunione Scientifiche Italiana, 1 (Pisa, 1839)*. *Annali di Fisica del Regno Lomb.-Ven.* **10**, 50.
- Newell, N. D. & Boyd, D. W. 1975. Parallel evolution in early Trigonian bivalves. *Bulletin of the American Museum of Natural History* **154**, 55–162.
- Owen, G. 1959. The ligament and digestive system in the taxodont bivalves. *Proceedings of the Malacological Society* **33**, 215–23.
- Owen, G., Trueman, E. R. & Yonge, C. M. 1953. The ligament in the Lamellibranchia. *Nature* **171**, 73–75.
- Powell, A. W. B. 1937. New species of marine Mollusca from New Zealand. *Discovery Report* **15**, 153–222.
- Scarlato, O. A. & Starobogatov, Y. I. 1983. Sistema dvustvorchatykh molliuskov nadotriada Septibranchia. [Classification of the bivalve molluscs of the Order Septibranchia.] In Likharev, I. M. (ed.): *Molliuski: sistematika, ekologii i zakonomernosti rasprostraneniia, Avtoferaty dokladov*, 7–13. [Molluscs: systematics, ecology and natural distribution. Abstracts of communications.] Sed'moe Vsesoiuznoe Soveshchaniepo Izucheniiu Molliuskov. [Seventh Soviet Conference on Molluscan Studies.] Zoologicheskii Institut, Leningrad: Akademiia Nauk SSSR. 262 pp.
- Smith, E. A. 1907. Mollusca – V. Lamellibranchiata. *National Antarctic Expedition 1901–1904 (Natural History, Zoology)* **2**, 1–7.
- Stanley, S. M. 1975a. Why clams have the shape they have: an experimental analysis of burrowing. *Paleobiology* **1**, 48–58.
- Stanley, S. M. 1975b. Adaptive themes in the evolution of the Bivalvia (Mollusca). *Annual Review of Earth and Planetary Sciences* **3**, 361–85.
- Stanley, S. M. 1977. Coadaptation in the Trigonidae, a remarkable family of burrowing bivalves. *Paleontology* **20**, 869–99.
- Stanley, S. M. 1978. Aspects of the adaptive morphology and evolution of the Trigonidae. *Philosophical Transactions of the Royal Society, London, B* **284**(1001), 247–58.
- Thiele, J. & Jaekel, S. G. A. 1931. Muscheln der Deutschen Tiefsee-Expedition. *Wissenschaftliche Ergebnisse der deutschen Tiefsee-Expedition auf dem Dampfer Valdivia 1898–1899* **21**, 161–268.
- Trueman, E. R. 1949. The ligament of *Tellina tenuis*. *Proceedings of the Zoological Society of London* **119**, 719–42.
- Waller, T. R. 1990. The evolution of ligament systems in the Bivalvia. In Morton, B. (ed.) *The Bivalvia* (Proceedings of a Memorial Symposium in Honour of Sir Charles Maurice Yonge (1899–1986) at the IXth International Malacological Congress, Edinburgh, 1986), 49–71. Hong Kong: Hong Kong University Press. 364 pp.
- Warton, D. I., Wright, I. J., Falster, D. S. & Westoby, M. 2006. Bivariate line fitting methods for allometry. *Biological Reviews of the Cambridge Philosophical Society* **81**(2), 259–91.
- Weaver, C. 1931. Paleontology of the Jurassic and Cretaceous of West Central Argentina. *Memoir of the University of Washington* **1**, 1–469.
- Wilbur, K. M. & Owen, G. 1964. Growth In Wilbur, K. M. & Yonge, C. M. (eds) *Physiology of Mollusca, Volume I*, 211–42. New York & London: Academic Press. 488 pp.
- Wilton, C. P. N. 1830. Memoir on the geology of the shore of the Severn, in the parish of Awre, Gloucestershire. *Quarterly Journal of Science, Literature and Arts* **Jan–June**, 64–73.
- Woodring, W. P. 1925. Miocene Mollusca from Bowden Jamaica, pelecypods and scaphopods. *Carnegie Institute, Washington, Publication* **366**, 1–564.
- Yonge, C. M. 1928. Structure and function of the organs of feeding and digestion in the Septibranchs, *Cuspidaria* and *Poromya*. *Philosophical Transactions of the Royal Society, London, B* **216**, 221–62.
- Yonge, C. M. 1939. The Protobranchiate Mollusca; a functional interpretation of their structure and evolution. *Philosophical Transactions of the Royal Society, London, B* **230**, 79–147.
- Yonge, C. M. 1978. Significance of the ligament in the classification of the Bivalvia. *Proceedings of the Royal Society, London, B* **202**, 231–48.
- Yonge, C. M. & Morton, B. 1980. Ligament and lithodesma in the Pandoracea and Poromyacea with a discussion on evolutionary history in the Anomalodesmata (Mollusca: Bivalvia). *Journal of Zoology* **191**, 263–92.
- Yonge, C. M. & Thompson, T. E. 1976. *Living marine Molluscs*. Glasgow: William Collins Sons & Co. Ltd. 288 pp.

MS received 15 May 2015. Accepted for publication 2 October 2016.

