

Determinants of Tissue PCO₂ in Shock and Sepsis: Relationship to the Microcirculation

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Introduction

The development of gastrointestinal tonometry was an important step in the monitoring of tissue dysoxia. It rapidly became a useful tool in basic research. In addition, and for the first time, a regional parameter could be used to detect and to treat hypoperfusion. From an experimental point of view, tonometry adequately tracks intramucosal acidosis [1], i.e., the increase in intramucosal-arterial PCO₂ difference (Δ PCO₂). Likewise, the increase in Δ PCO₂ is better than other systemic and intestinal variables to show tissue hypoperfusion in normal volunteers [2] and in experimental models [3]. Intramucosal acidosis is a sensitive predictor of gastric [4] and colonic mucosal ischemia [5]. Furthermore, gastric tonometry is an insightful predictor of outcome. This usefulness has been shown in postoperative [6], critically ill [7], septic [8] and shock [9] patients. Gastric tonometry might also be used to assess the effect of vasoactive drugs [10, 11]. Finally, intramucosal pH (pHi) has been evaluated as a guide for resuscitation. Gutierrez et al. [12] demonstrated in a randomized controlled trial that pHi-guided therapy could decrease mortality in critically ill patients.

Despite having been the only clinically available approach to detect tissue hypoperfusion for many years and despite the scientific evidence supporting its usefulness, gastrointestinal tonometry is not commonly used. Various reasons may explain this issue, including that saline tonometry has poor reproducibility [13], although this was improved by the introduction of air tonometry [14]. Sublingual capnometry remains an attractive approach [15], but this technique has not yet been adequately validated.

Another source of uncertainty lies in the true significance of Δ PCO₂ elevation. In the last few years, new evidence has given a better understanding of the mechanisms underlying intramucosal acidosis. In this chapter, we will discuss the determinants of tissue and venous PCO₂ in shock and sepsis and their relationship to microcirculatory perfusion.

Mechanisms of Increase in Venous and Tissue PCO₂: The Basics

Increased mucosal intestinal PCO₂ has been mainly used to detect tissue dysoxia, the condition in which oxygen delivery (DO₂) can no longer sustain oxygen consumption (VO₂) [16]. Twenty years ago, Grum et al. [17] evaluated the adequacy of gut oxygenation by the tonometric measurement of pHi, during DO₂ reductions secondary to ischemia, hypoxemia, or a combination of both. pHi only decreased after crit-

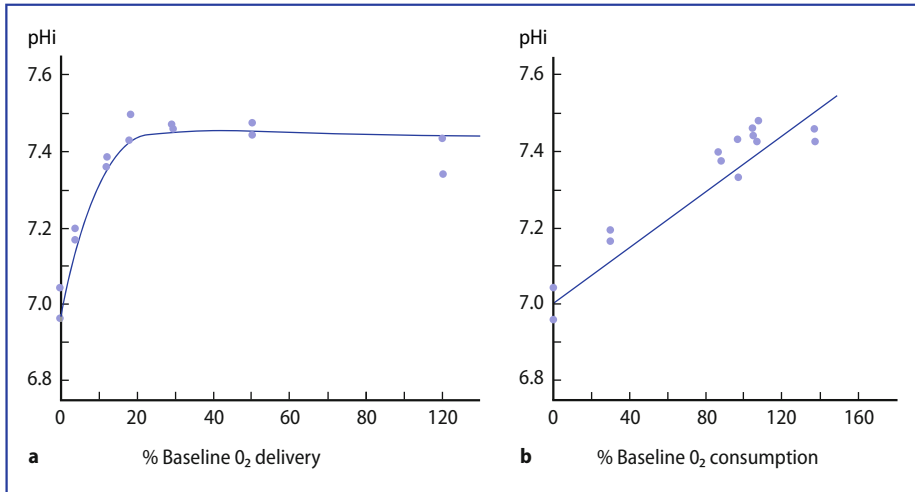


Fig. 1a. Intestinal intramucosal pH (pHi) as a function of oxygen (O₂) delivery. **b** Intestinal intramucosal pH as a function of intestinal O₂ consumption. Reductions of intramucosal pH are only present with critical reductions of O₂ delivery. From [17] with permission.

ical reductions of DO₂. Consequently, changes in VO₂ and pHi were closely correlated (**Fig. 1**). The authors concluded that pHi appears to be a sensitive indicator of tissue oxygenation, because it mirrors tissue VO₂. Nevertheless, critical DO₂ was only reached in ischemic experiments. In pure hypoxemic experiments, neither pHi nor VO₂ decreased.

Theoretically, PCO₂ can increase in the intestinal lumen by two mechanisms [18]: First, by bicarbonate buffering of the protons generated during the breakdown of high-energy phosphates and strong acids, in which case increased PCO₂ would represent tissue dysoxia; alternatively, PCO₂ can increase due to hypoperfusion and decreased washout of CO₂. The Fick Equation applied to CO₂, states that CO₂ production (VCO₂) is the product of cardiac output and venoarterial CO₂ content difference. Consequently, decreases in blood flow result in venous and tissue hypercarbia, regardless of the lack of change in VCO₂.

Trying to solve this controversy, Schlichtig and Bowles [18] presented evidence supporting intramucosal PCO₂ as a marker of dysoxia in extreme hypoperfusion when VO₂ decreases. In a dog model of cardiac tamponade, these authors demonstrated that below critical DO₂, mucosal PCO₂ increases because of anaerobic VCO₂. This conclusion was drawn using the Dill nomogram, which can, theoretically, detect anaerobic VCO₂ from the comparison of the measured (%HbO_{2v}) vs. calculated (%HbO_{2v}^{DILL}) venous oxyhemoglobin, within a given value of venous PCO₂. Since venous PCO₂ is considered representative of tissue PCO₂, the authors made the calculation with its intestinal equivalent, intramucosal PCO₂. Similar values of measured (%HbO_{2v}) vs. calculated (%HbO_{2v}^{DILL}) venous oxyhemoglobin would represent aerobic VCO₂. If %HbO_{2v}^{DILL} is lower than measured %HbO_{2v}, anaerobic VCO₂ is then assumed. Using this approach, we identified an anaerobic source of gut intramucosal CO₂ during moderate hemorrhage [3]. Our %HbO_{2v}^{DILL} values obtained from gastric, jejunal, and ileal mucosal PCO₂ decreased markedly during ischemia, indicating the presence of anaerobias (**Fig. 2**). Notwithstanding the original contribu-

Fig. 2. Measured venous oxygen saturation (%HbO_{2v}) (□) and venous oxygen saturation calculated from gut tissue PCO₂ (%HbO_{2v}^{DILL}) (●) as a function of superior mesenteric artery blood flow. Since %HbO_{2v}^{DILL} is lower than measured %HbO_{2v}, anaerobic production of CO₂ may be assumed. From [3] with permission.

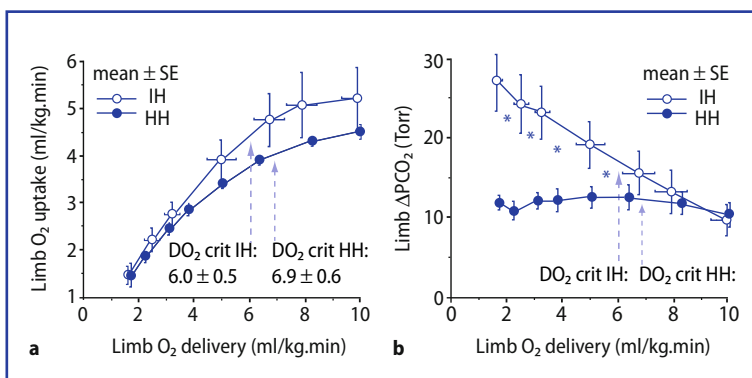
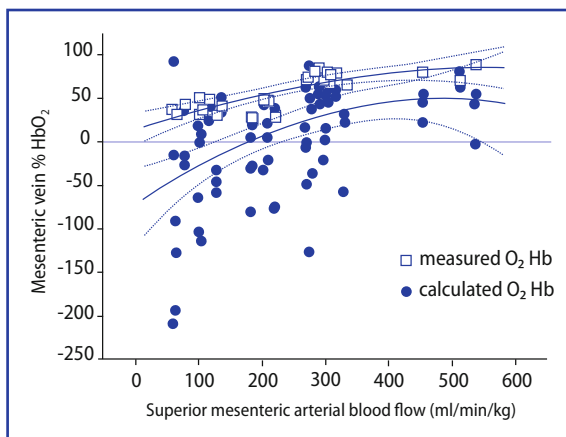


Fig. 3. a Hindlimb oxygen uptake as a function of limb oxygen delivery (DO₂) for ischemic hypoxia (IH) and hypoxic hypoxia (HH). There was no statistically significant difference at any DO₂. Critical DO₂ (DO₂crit) was not different in IH and HH. **b** Hindlimb venoarterial PCO₂ difference as a function of limb DO₂ for IH and HH. Despite similar degrees of tissue dysoxia, venoarterial PCO₂ difference remained constant in the HH group and increased more than twofold in the IH group. From [20] with permission.

tion of Schlichtig and Bowles [18] to the analysis of these topics, the use of low flow to produce critical DO₂ and decreased VO₂ may act as a potential confounder, given the impossibility of dissociating tissue dysoxia from hypoperfusion [19].

Vallet et al. [20] explored this issue by measuring venous PCO₂ in isolated dog hindlimb preparations subjected to comparable decreases in DO₂, produced by two mechanisms. In one group, blood flow was progressively decreased (ischemic hypoxia), whereas in the other, arterial PO₂ was lowered at constant perfusion flow (hypoxic hypoxia). Both groups experienced similar declines in DO₂ and VO₂, implying similar degrees of tissue dysoxia. The venoarterial PCO₂ difference, however, remained constant in the hypoxic hypoxia group, and increased more than twofold in the ischemic hypoxia group. The authors concluded that flow is the major determinant of venoarterial PCO₂ difference, not tissue dysoxia [20] (Fig. 3).

Nevière et al. assessed a similar hypothesis in pigs, comparing the effects of reduced inspired oxygen fraction (FiO₂) and decreased blood flow measured with

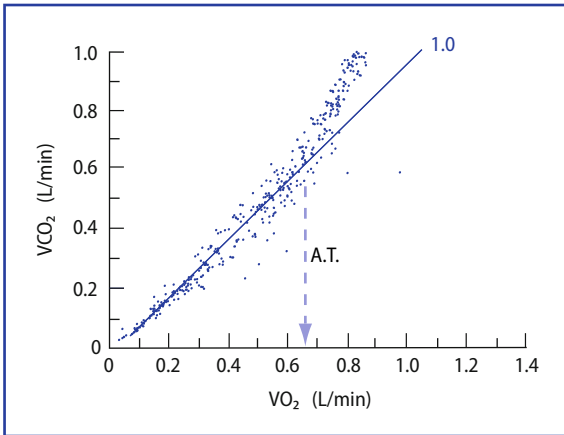


Fig. 4. Plot of carbon dioxide production (VCO_2) as a function of oxygen uptake (VO_2) in response to incremental work rate during a test of exercise. Line with slope 1.0 is the approximate mean of data points up to the point where VCO_2 breaks away and increases rapidly. When data points rise more steeply than a slope of 1.0 is theoretically the VCO_2 at which HCO_3^- starts buffering lactic acid (anaerobic threshold). From [22] with permission.

laser-Doppler [21]. In ischemic hypoxia, ΔPCO_2 rose to 60 mmHg. In hypoxic hypoxia, in which mucosal blood flow was maintained constant, ΔPCO_2 increase to 30 mmHg only with the lowest FiO_2 (0.06). The authors concluded that intramucosal PCO_2 elevation in hypoxic hypoxia denotes local CO_2 generation. Some flow heterogeneity could, however, have been present in their experiments that was not assessed by laser-Doppler, a method that only tracks global microvascular changes. In addition, in the two preceding steps of FiO_2 reduction, VO_2/DO_2 dependency had been reached, and ΔPCO_2 remained unchanged.

From a physiologic point of view, it is difficult to understand how VCO_2 might increase during oxygen supply dependency. During progressive exercise, there are corresponding increases in VO_2 and VCO_2 [22]. The slope of the VCO_2/VO_2 relationship is the respiratory quotient. When the exercise reaches the anaerobic threshold, there is an excess of VCO_2 to VO_2 due to the appearance of anaerobic VCO_2 from the bicarbonate buffering of lactic acid (Fig. 4). In this condition, both VCO_2 and respiratory quotient increase.

In the other extreme of physiology, during oxygen supply dependency, the respiratory quotient also increases [23]. This increase, however, occurs in the context of the reduction of total VCO_2 (Fig. 5). Anaerobic VCO_2 appears, but total VCO_2 decreases.

We further explored this issue in another model of hypoxic hypoxia [24]. In these experiments, venous and tissue PCO_2 increased during ischemic hypoxia, but not during hypoxic hypoxia. Therefore, ΔPCO_2 was unable to show the presence of tissue dysoxia during hypoxic hypoxia, in which blood flow is preserved (Fig. 6a). To confirm that blood flow is the main determinant of ΔPCO_2 , we studied these relationships in another model of tissue dysoxia without hypoperfusion, anemic hypoxia [25] (Fig. 6b). We compared the effects of progressive bleeding to those of isovolemic exchange of blood with dextran. Our goal was to evaluate the behavior of CO_2 gradients as a function of systemic and intestinal blood flow, and also the other determinants, VCO_2 and the CO_2 Hb dissociation curve. Tissue-arterial and venoarterial PCO_2 failed to reflect the dependence of VO_2 on DO_2 . Nevertheless, these gradients increased by a few mmHg (Figs. 6 and 7). Conversely, however, venoarterial CO_2 content differences decreased. This apparent paradox might be explained by changes in the CO_2 Hb dissociation curve induced by anemic hypoxia. The other determinant of PCO_2 differences, the VCO_2 remained unchanged, both at systemic and intestinal

Fig. 5. Effects of progressive bleeding on oxygen consumption (VO₂), carbon dioxide production (VCO₂), and respiratory quotient. Panel **a**. After critical reductions in DO₂, VO₂ and VCO₂ decreased. Panel **b**. After critical reductions in DO₂, respiratory quotient increased [23].

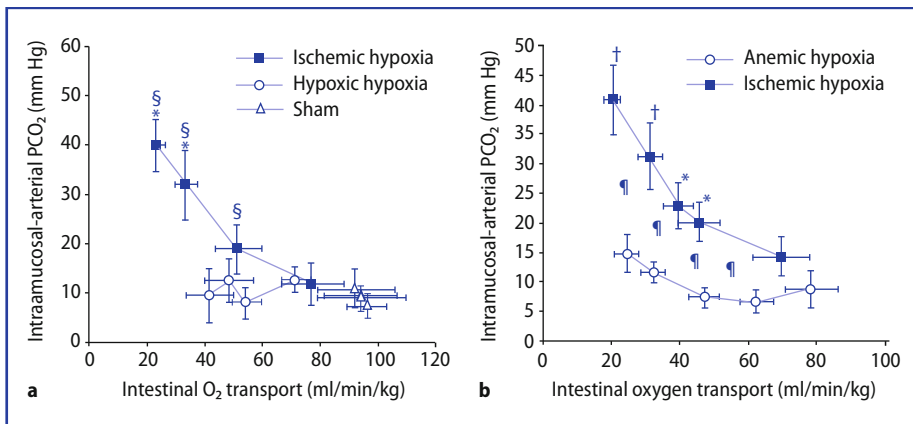
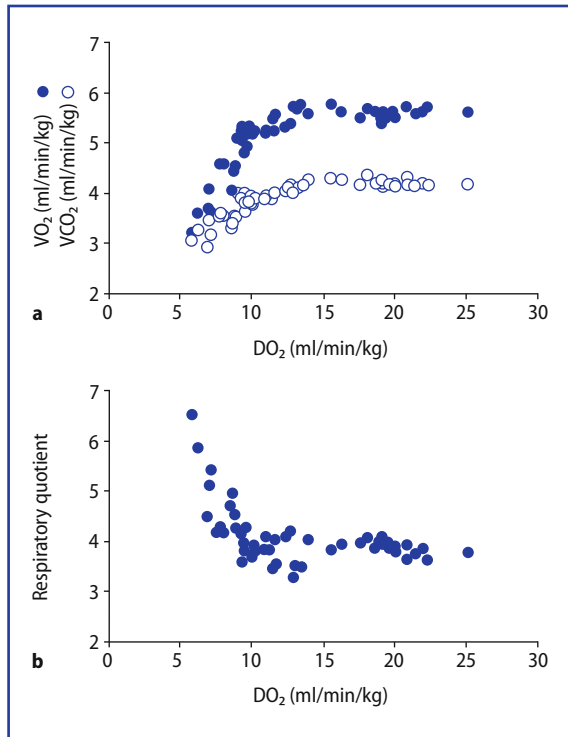


Fig. 6. a Ileal intramucosal-arterial PCO₂ difference (Δ PCO₂) as a function of intestinal oxygen transport in hypoxic and ischemic hypoxia. From [24] with permission. **b** Δ PCO₂ as a function of intestinal oxygen transport in anemic and ischemic hypoxia. From [25] with permission. In hypoxic and anemic hypoxia, Δ PCO₂ fails to reflect tissue dysoxia.

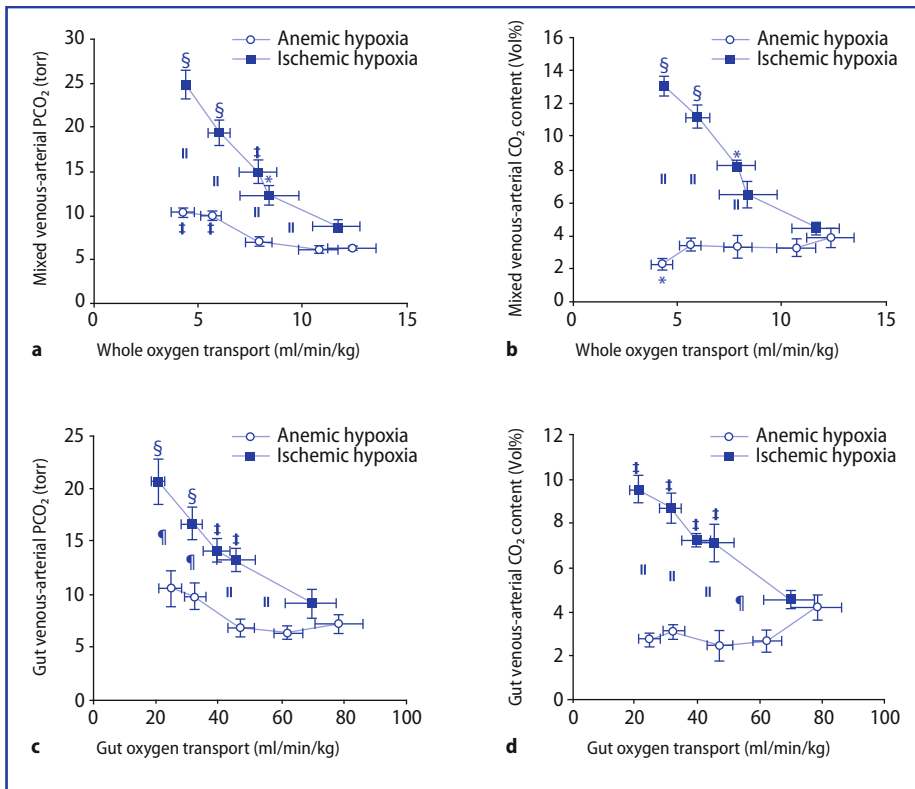


Fig. 7. **a** Mixed venous-arterial PCO₂ difference as a function of systemic oxygen transport (DO₂) in ischemic and anemic hypoxia. **b** Mixed venous-arterial CO₂ content difference as a function of systemic DO₂ in ischemic and anemic hypoxia. **c** Mesenteric venous-arterial PCO₂ difference as a function of intestinal DO₂ in ischemic and anemic hypoxia. **d** Mesenteric venous-arterial CO₂ content difference as a function of intestinal DO₂ in ischemic and anemic hypoxia. Differences were higher in ischemic than in anemic hypoxia. Venous-arterial PCO₂ differences slightly increased while venous-arterial CO₂ content differences decreased in anemic hypoxia, implying changes in the CO₂Hb dissociation curve. From [25] with permission.

levels. The systemic and intestinal respiratory quotient, however, increased because of VO₂ reductions.

In summary, our results [24, 25], together with those of Vallet et al. [20], support the concept that increases in tissue-arterial and venous-arterial PCO₂ gradients reflect only microcirculatory stagnation, not tissue dysoxia. Tissue and venous PCO₂ are insensitive markers of dysoxia and merely indicate hypoperfusion. These experimental findings were confirmed by a mathematical model [26]. Gutierrez developed a two-compartment mass transport model of tissue CO₂ exchange for hypoxic hypoxia, to examine the relative contribution of blood flow and cellular dysoxia to the increases in tissue and venous PCO₂. The model assumed perfectly mixed homogeneous conditions, steady-state equilibrium, and VCO₂ occurring exclusively at the tissues. The results of the model supported the idea that changes in tissue and venous blood CO₂ concentrations during dysoxia reflect primarily alterations in vascular perfusion, and not shortage of energy supply.

Intramucosal Acidosis in Sepsis

Beyond the previous discussion, intramucosal acidosis is a common finding in clinical and experimental sepsis, conditions in which cardiac output is usually normal or increased. In resuscitated endotoxemic pigs, VanderMeer et al. found that intramucosal acidosis developed despite preserved mucosal oxygenation and blood flow measured only at the mucosa [27]. The underlying mechanism was attributed to metabolic disturbances, and led to the concept of “cytopathic hypoxia” [28]. Nevertheless, an important shortcoming of that study was the use of laser-Doppler flowmetry to measure tissue perfusion and the lack of measurement of tissue oxygenation at the serosal side of the intestines, an important source of gut CO₂ [31].

On the other hand, Vallet et al. studied dogs challenged with endotoxin, and then resuscitated them to normalize oxygen transport [29]. Intestinal VO₂ and mucosal PO₂ and pH, however, remained low. The authors ascribed these findings to blood flow redistribution from the mucosa toward the muscular layer. Nevertheless, Revelly et al. [30] described an inverse redistribution, with increased mucosal and decreased muscular blood flow, using dyed microspheres in endotoxemic pigs [30]. Paradoxically, pHi was inversely correlated with mucosal flow, though positively correlated with muscular perfusion. The authors concluded that intramucosal acidosis was not explained by mucosal hypoperfusion [30]. Siegemund et al. showed, in a similar model, reductions in mucosal and serosal microvascular PO₂, and an increase in ΔPCO₂ [31]. Fluid resuscitation normalized mucosal PO₂ but serosal PO₂ and ΔPCO₂ remained altered. Inhibition of inducible nitric oxide (NO) however restored serosal PO₂ and also ΔPCO₂ thereby identifying the source of intraluminal CO₂ measured in their model.

Conversely, Tugtekin et al. [32], in a porcine model of 24-hour endotoxin infusion, showed an association between intramucosal acidosis and severe hypoperfusion in ileal villi. In this study, about half of the evaluated villi were heterogeneously- or non-perfused, despite normal portal blood flow. Creteur et al. described, in septic patients, a correlation between sublingual ΔPCO₂ and microcirculatory blood flow [33]. In agreement with their results, results from our laboratory showed that endotoxic shock in sheep was associated with sublingual and intestinal microcirculatory alterations and intramucosal acidosis [34]. Fluid resuscitation normalized systemic and intestinal oxygen transport, as well as sublingual and intestinal serosal microcirculation. Nevertheless, a reduced number of perfused intestinal villi and increased ΔPCO₂ persisted. This led us to conclude that intramucosal acidosis was related to a persistent decrease in the mucosal microvascular flow index and a reduced number of perfused intestinal villi [34] (Fig. 8).

There are other studies further supporting the hypothesis that, in endotoxemia, changes in perfusion and not tissue dysoxia determine ΔPCO₂. We randomized endotoxemic sheep to saline solution resuscitation to maintain blood flow at baseline values or to increase it by 50%. Increased perfusion prevented intramucosal acidosis, though metabolic acidosis continued due to increased anion gap [35]. Similarly, in endotoxemic sheep, the administration of levosimendan, an inotropic and vasodilator drug, precluded increases in ΔPCO₂ but hyperlactatemia was exacerbated [36] or unaffected [37]. The findings of these studies suggest that intramucosal acidosis is mainly related to local hypoperfusion and that metabolic disorders depend on a cellular mechanism which is unresponsive to changes in blood flow.

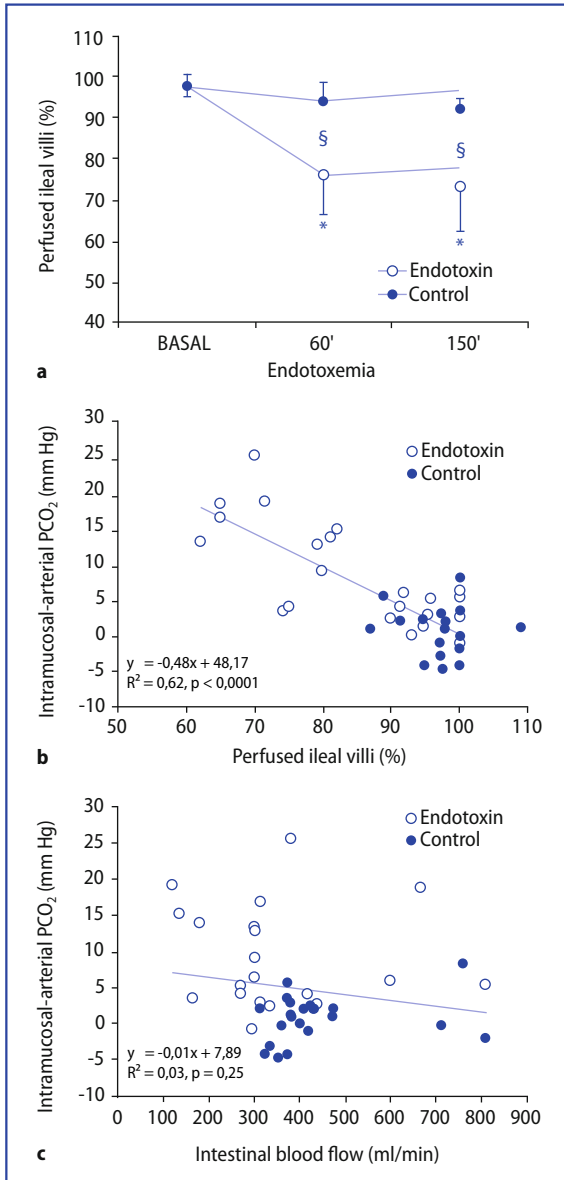


Fig. 8. Effects of endotoxic shock and resuscitation on the percentage of perfused ileal villi and intramucosal-arterial PCO₂ difference (ΔPCO₂). Endotoxic shock decreased the perfused intestinal villi and fluid resuscitation was unable to restore villus perfusion (a). ΔPCO₂ was correlated with perfused intestinal villi (b) but not with superior mesenteric artery blood flow (c). From [34] with permission.

Conclusion

Venoarterial and tissue-arterial PCO₂ gradients are the result of interactions in aerobic and anaerobic VCO₂, CO₂ dissociation curve, and blood flow to tissues. During VO₂/DO₂ dependency, opposite changes in aerobic and anaerobic VCO₂ occur. Aerobic VCO₂ decreases as a consequence of failing aerobic metabolism, but, at the same time, anaerobic VCO₂ starts due to bicarbonate buffering of protons derived from

fixed acids. Total VCO₂ might not increase, as in our experiments. But as VO₂ falls, there is an increase in the respiratory quotient [27]. The relative increment of VCO₂ with respect to VO₂ can only cause venous and tissue hypercarbia during tissue hypoperfusion, in which CO₂ removal is reduced. These conditions can be present despite preserved systemic and regional blood flow.

Notwithstanding the fact that ΔPCO₂ is not a marker of dysoxia but of tissue perfusion, it remains a very useful clinical and experimental monitoring tool, particularly in clinical situations such as sepsis in which cardiac output is increased while microcirculatory flow can be impaired.

References

1. Antonsson JB, Boyle CC 3rd, Kruihoff KL, et al (1990) Validation of tonometric measurement of gut intramural pH during endotoxemia and mesenteric occlusion in pigs. *Am J Physiol* 259: G519-G523
2. Hamilton-Davies C, Mythen MG, Salmon JB, Jacobson D, Shukla A, Webb AR (1997) Comparison of commonly used clinical indicators of hypovolaemia with gastrointestinal tonometry. *Intensive Care Med* 23: 276–281
3. Dubin A, Estensoro E, Murias G, et al (2001) Effects of hemorrhage on gastrointestinal oxygenation. *Intensive Care Med* 27: 1931–1936
4. Fiddian-Green R, McGough E, Pittenger G, Rothman E (1983) Predictive value of intramural pH and other risk factors for massive bleeding from stress ulceration. *Gastroenterology* 85: 613–620
5. Schiedler MG, Cutler BS, Fiddian-Green R (1987) Sigmoid intramural pH for prediction of ischemic colitis during aortic surgery. A comparison with risk factors and inferior mesenteric artery stump pressures. *Arch Surg* 122: 881–886
6. Mythen M, Webb A (1994) Intra-operative gut mucosal hypoperfusion is associated with increased post-operative complications and cost. *Intensive Care Med* 20: 99–104
7. Doglio G, Pusajo J, Egurrola M, et al (1991) Gastric mucosal pH as a prognostic index of mortality in critically ill patients. *Crit Care Med* 19: 1037–1040
8. Friedman G, Berlot G, Kahn R, Vincent JL (1995) Combined measurements of blood lactate concentrations and gastric intramucosal pH in patients with severe sepsis. *Crit Care Med* 23: 1184–1193
9. Maynard N, Bihari D, Beale R, et al (1993) Assessment of splanchnic oxygenation by gastric tonometry in patients with acute circulatory failure. *JAMA* 270: 1203–1210
10. Gutierrez G, Clark C, Brown SD, Price K, Ortiz L, Nelson C (1994) Effect of dobutamine on oxygen consumption and gastric mucosal pH in septic patients. *Am J Respir Crit Care Med* 150: 324–329
11. Nevière R, Mathieu D, Chagnon JL, Lebleu N, Wattel F (1996) The contrasting effects of dobutamine and dopamine on gastric mucosal perfusion in septic patients. *Am J Respir Crit Care Med* 154: 1684–1688
12. Gutierrez G, Palizas F, Doglio G, et al (1992) Gastric intramucosal pH as a therapeutic index of tissue oxygenation in critically ill patients. *Lancet* 339: 195–199
13. Oud L, Kruse J (1996) Poor in vivo reproducibility of gastric intramucosal pH determined by saline-filled balloon tonometry. *J Crit Care* 11: 144–150
14. Taylor D, Gutierrez G, Clark C, Hainley S (1997) Measurement of gastric mucosal carbon dioxide tension by saline and air tonometry. *J Crit Care* 12: 208–213
15. Toledo Maciel A, Creteur J, Vincent JL (2004) Tissue capnometry: does the answer lie under the tongue? *Intensive Care Med* 30, 2157–2165
16. Honig C, Connert R, Gayeski T, Brooks G (1990) Defining hypoxia: a systems view of VO₂, glycolysis, energetics, and intracellular PO₂. *J Appl Physiol* 68: 833–842
17. Grum C, Fiddian-Green R, Pittenger G, Grant B, Rothman E, Dantzker D (1984) Adequacy of tissue oxygenation in intact dog intestine. *J Appl Physiol* 56: 1065–1069
18. Schlichtig R, Bowles S (1994) Distinguishing between aerobic and anaerobic appearance of dissolved CO₂ in intestine during low flow. *J Appl Physiol* 76: 2443–2451

19. Vallet B, Tavernier B, Lund N (2000) Assessment of tissue oxygenation in the critically ill. In: Vincent JL (ed) Yearbook of Intensive Care and Emergency Medicine. Springer-Verlag, Heidelberg, pp 715–725
20. Vallet B, Teboul JL, Cain S, Curtis S (2000) Venous-arterial CO₂ difference during regional ischemic or hypoxic hypoxia. *J Appl Physiol* 89: 1317–1321
21. Nevière R, Chagnon JL, Teboul JL, Vallet B, Wattel F (2002) Small intestine intramucosal PCO₂ and microvascular blood flow during hypoxic and ischemic hypoxia. *Crit Care Med* 30, 379–384
22. Wasserman K, Beaver WL, Whipp BJ (1990) Gas exchange theory and the lactic acidosis (anaerobic) threshold. *Circulation* 81 (1 Suppl):II14–30
23. Dubin A, Murias G, Estenssoro E, et al (2000) End-tidal CO₂ pressure determinants during hemorrhagic shock. *Intensive Care Med* 26: 1619–1623
24. Dubin A, Murias G, Estenssoro E, et al (2002) Intramucosal-arterial PCO₂ gap fails to reflect intestinal dysoxia in hypoxic hypoxia. *Crit Care* 6: 514–520
25. Dubin A, Estenssoro E, Murias G, et al (2004) Intramucosal-arterial PCO₂ gradient does not reflect intestinal dysoxia in anemic hypoxia. *J Trauma* 57: 1211–1217
26. Gutierrez G (2004) A mathematical model of tissue-blood carbon dioxide exchange during hypoxia. *Am J Respir Crit Care Med* 169: 525–533
27. VanderMeer T, Wang H, Fink M (1995) Endotoxemia causes ileal mucosal acidosis in the absence of mucosal hypoxia in a normodynamic porcine model of septic shock. *Crit Care Med* 23: 1217–1226
28. Fink M (2002) Bench-to-bedside review: Cytopathic hypoxia. *Crit Care* 6: 491–499
29. Vallet B, Lund N, Curtis S, Kelly D, Cain S (1994) Gut and muscle tissue PO₂ in endotoxemic dogs during shock and resuscitation. *J Appl Physiol* 76: 793–800
30. Revelly J-P, Ayuse T, Brienza N, Fessler H, Robotham J (1996) Endotoxic shock alters distribution of blood flow within the intestinal wall. *Crit Care Med* 24: 1345–1351
31. Siegemund M, van Bommel J, Schwarte L, et al (2005) Inducible nitric oxide synthase inhibition improves intestinal microcirculatory oxygenation and CO₂ balance during endotoxemia in pigs. *Intensive Care Med* 31: 985–992
32. Tugtekin IF, Radermacher P, Theisen M, et al (2001) Increased ileal-mucosal-arterial PCO₂ gap is associated with impaired villus microcirculation in endotoxic pigs. *Intensive Care Med* 27: 757–766
33. Creteur J, De Backer D, Sakr Y, Koch M, Vincent JL (2006) Sublingual capnometry tracks microcirculatory changes in septic patients. *Intensive Care Med* 32: 516–523
34. Dubin A, Kanoore Edul V, Murias G, et al (2008) Persistent villi hypoperfusion explains intramucosal acidosis in sheep endotoxemia. *Crit Care Med* 36: 535–542
35. Dubin A, Murias G, Maskin B, et al (2005) Increased blood flow prevents intramucosal acidosis in sheep endotoxemia: a controlled study. *Crit Care* 9: R66–73
36. Dubin A, Maskin B, Murias G, et al (2006) Effects of levosimendan in normodynamic endotoxaemia: a controlled experimental study. *Resuscitation* 69: 277–286
37. Dubin A, Murias G, Sottile J; et al (2007) Effects of levosimendan and dobutamine in experimental acute endotoxemia: A preliminary controlled study. *Intensive Care Med* 33: 485–494