

Simuliid Blackflies (Diptera: Simuliidae) and Ceratopogonid Midges (Diptera: Ceratopogonidae) as Vectors of *Mansonella ozzardi* (Nematoda: Onchocercidae) in Northern Argentina

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Mansonella ozzardi, a relatively non pathogenic filarial parasite of man in Latin America, is transmitted by either ceratopogonid midges or simuliid blackflies. In the only known focus of the disease in north-western Argentina the vectors have never been incriminated. This study investigated the potential vectors of *M. ozzardi* in this area. The only anthropophilic species of these Diptera families biting man at the time of the investigation were *Simulium exiguum*, *S. dinellii*, *Culicoides lahillei* and *C. paraensis*. Using experimentally infected flies *S. exiguum* and both species of *Culicoides* allowed full development of microfilariae to the infective stage, with *C. lahillei* being a more competent host than *S. exiguum*. Based on these data, biting rates and natural infectivity rates it is probable that at the beginning of the wet season *C. lahillei* is the main vector of *M. ozzardi* and both *C. paraensis* and *S. exiguum* secondary vectors. Additionally, it was found that a single dose of ivermectin was ineffectual in eradicating *M. ozzardi* from infected individuals in this area.

Key words: *Mansonella ozzardi* - Simuliidae - Ceratopogonidae - *Simulium exiguum* - *Simulium dinellii* - *Culicoides lahillei* - *Culicoides paraensis* - Argentina

Mansonella ozzardi is a filarial parasite first described from Guyana (then British Guiana) at the end of the last century and now known to occur in Central America in the Yucatan Peninsula of Mexico through to northern Argentina in South America, as well as on various Caribbean islands. It is relatively non pathogenic, its main importance lying in its morphological similarity to the highly pathogenic *Onchocerca volvulus*. This is especially relevant in Brazil where the two species are sympatric in parts of the Amazônia onchocerciasis focus. Attempts at controlling mansonelliasis have been few, but recent reports indicate that ivermectin may be effective as a microfilaricide. Various anthropophilic Diptera were examined before it was shown that *M. ozzardi* is transmitted by *Culicoides furens* and possibly *C. paraensis* in the Caribbean Island of St Vincent (reviewed by Buckley 1934, Shelley & Shelley 1976). Later, simuliid species

were incriminated as vectors in South America. This gave rise to the hypothesis that two races of *M. ozzardi* exist: one in the Caribbean islands and Mexico transmitted by ceratopogonid species, and the other transmitted by simuliid species in Panama and South America (Nelson & Pester 1962). Currently, only *Culicoides* species are known to transmit in the Caribbean islands but both *Simulium* and *Culicoides* species are vectors in Central and South America (Table I), but no comprehensive studies have been carried out on the transmission of *M. ozzardi* and the relative competence of its vectors.

Several studies have been made on *M. ozzardi* in Argentina and these are reviewed in Romaña and Wygodzinsky (1950). Since then several reports have been published on communities in Argentina that are infected with this parasite (Undiano 1971, Jörg 1983, Minoprio et al. 1988, Remondegui et al. 1988, Taranto & Castelli 1988). These studies indicate that mansonelliasis is confined to the provinces of Tucumán and Jujuy to the north of the country and affects mainly subsistence farming communities, largely formed in the past by the southern dispersal of indians from southern Bolivia. It has been shown that the parasite is relatively non pathogenic, although treatment with ivermectin has been carried out. Attempts at establishing the identity of the vector of *M. ozzardi* in Argentina have failed. Buckley (1934) and

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TABLE I
 Vectors of *Mansonella ozzardi* in Latin America

Central America and Caribbean	Ceratopogonidae	Simuliidae	References
Haiti	<i>Culicoides furens</i> , <i>C. barbosai</i> , <i>Leptoconops bequaerti</i>		Lowrie and Raccurt 1981,1984 Lowrie et al. 1983
Mexico	<i>C. furens</i>		Biagi et al. 1958
Panama		<i>Simulium sanguineum</i>	Petersen et al. 1984
St Vincent	<i>C. furens</i> , <i>C. paraensis?</i>		Buckley 1934
Trinidad	<i>C. phlebotomus</i>		Nelson and Davies 1976
South America			
Argentina	<i>C. paraensis?</i>		Romaña and Wygodzinsky 1950
Brazil		<i>S. amazonicum</i> , <i>S. argentiscutum</i> <i>S. oyapockense s.l.</i> or <i>S. roraimense</i>	Moraes et al. 1985 Shelley et al. 1980
Colombia	<i>C. insinuatus</i> , <i>C. caprilesi?</i>	<i>S. amazonicum</i> , <i>S. argentiscutum</i> <i>S. oyapockense s.l.</i> <i>S. sanguineum</i>	Tidwell and Tidwell 1982 Tidwell et al. 1980
Guyana		<i>S. oyapockense s.l.</i>	Nathan et al. 1982
Surinam	<i>C. guttatus?</i>		Bruijning 1957
Venezuela		<i>S. oyapockense s.l.</i>	Yarzabal et al. 1985

Romaña and Wygodzinsky (1950) cite previous experimental infection studies carried out in Argentina showing that *Cimex* (Hemiptera, Cimicidae) and *Triatoma* (Hemiptera, Reduviidae) bugs were not hosts, that microfilariae developed to the L1 stage in *Anopheles tarsimaculatus*, *A. albittarsis* and *Aedes aegypti* but not in *A. pseudopunctipennis*, *A. argyritarsis* and *C. quinquefasciatus* (Diptera, Culicidae). Romaña and Wygodzinsky (1950) showed that no development occurred in simuliids that had fed on *M. ozzardi* carriers, but that development did occur to the L2 stage in *C. paraensis*. Development to the L3 stage did not continue because of premature mortality of flies.

The objective of the current work was to discover the vector species of *M. ozzardi* in the mansonelliasis focus in northern Argentina.

MATERIALS AND METHODS

Study area - The work was carried out in January 1998 at the beginning of the wet season in subtropical cloud forest (Yungas) in the mountain range Quebrada de Humahuaca, to the east of the Andean foothills and north of San Salvador de

Jujuy in Jujuy Province in north-west Argentina. The sparse population (300) in this area, drained by the River Candelaria and its tributaries, is involved in subsistence agriculture and timber extraction; in the lower lying areas nearer the town of Fraile Pintado orange groves and sugar plantations provide the main agricultural activities. Laboratory facilities were used in the hospital in the town of Fraile Pintado and the school at Aforo (previously Arrayanal, 23°54'S65°02'W, altitude 900 m), some 50 km to the west of this town on the confluence of the River Candelaria and Arroyo Normenta, was used as a base for field work. This was centred on riverine communities (called Finca Normenta) of mainly local subsistence farmers living farther up the Arroyo Normenta.

Selection of volunteers - Remondegui et al. (1988) carried out a survey of mansonelliasis and its potential vectors in the Finca Normenta area ten years ago and found prevalence rates in villages varying between 25-75%. In November 1995 a survey was made in the same area by staff from the hospital at Fraile Pintado to register the names of individuals infected with *M. ozzardi*, followed in December 1995 by a single treatment of infected

individuals using ivermectin (pers. comm. Dr G Safarov). This register of individuals infected with mansonelliasis with information on their treatment was used as a basis for selecting people for the present study. A lancet was used to puncture the fourth finger of each individual and a drop of blood used to make a thick film on a microscope slide. The film was de-haemaglobinized, fixed in 80% methanol, stained with Giemsa and then examined for microfilariae.

Prospection for simuliid and ceratopogonid species - Fifty years ago Romaña and Wygodzinsky (1950) surveyed an area in Tucumán Province to the north of our study area and considered the following anthropophilic species to be potential vectors of *M. ozzardi*: *S. dinellii*, *S. jujuyense*, *C. paraensis* and *C. lahillei* (as *C. debilipalpis* - now considered a junior synonym of *C. lahillei*). Previous prospection of the Finca Normenta area by Remondegui et al. (1988) in the dry season (pers. comm. Dr G Safarov) revealed the presence of three simuliid species (two unnamed) of which *S. dinellii* was predominant and 16 species of ceratopogonids (all unnamed). In the present survey immature stages of Simuliidae were collected from the rivers Candelaria and Normenta for several hours on several days at the beginning of the study. Larvae and immature pupae were preserved in 80% ethanol, as were adults that emerged from mature pupae kept individually in tubes containing moistened paper. Linked reared adults were also micropinned and, together with their pupal exuviae, preserved in a small phial of glycerine attached to staging pins. Adult Simuliidae and Ceratopogonidae were also collected for short periods over several days biting ourselves, horses and cattle in order to establish which were the anthropophilic species in the area. Specimens of *C. lahillei* were later dissected for filariae. Identification of simuliid species was carried out by Prof. S Coscarón and the ceratopogonid species by Dr G Spinelli of the La Plata Museum. Voucher specimens have been deposited in the La Plata Museum and the Natural History Museum, London.

Infection of flies - In order to establish the hosts of *M. ozzardi* samples of local biting flies were experimentally fed with the parasite to observe any subsequent development and other flies were dissected for natural infections with filariae.

The volunteer used for experimental infection studies was a highly infected 74 years old male, who has lived in the area all his life. He was positive in the 1995 survey, was treated with ivermectin then, but was still highly positive in the present survey. Simuliids and ceratopogonids were allowed to feed to repletion on the volunteer's legs and a small proportion of these flies had their ab-

domens removed and the blood in their stomachs smeared on a microscope slide for counting and identification of microfilariae using the technique given above for thick films. The rest were maintained alive for up to nine days in individual tubes containing filter paper and a cotton wool wick soaked in 20% boiled sucrose solution. Tubes were kept in the dark in an insulated container in which the temperature varied from 21-30°C. The tubes were examined daily for dead flies, which were stored in 80% ethanol and all living flies nine days after experimental infection were sacrificed and similarly stored. The cotton wool wicks were changed every two days. Preserved flies were later examined in the laboratory for filariae using the following method. Flies were removed from ethanol and placed in tap water over night for re-hydration. Each fly was then placed in a drop of tap water and divided into head, thorax and abdomen. These parts were then added to a drop of Mayer's haemalum for 30 min, transferred to a drop of tap water to remove excess stain and then placed separately in three drops of glycerine on a microscope slide. The head, thorax and abdomen were then dissected and parasites removed from each site were mounted in Berlese mountant for identification using morphological characters described in Shelley et al. (1980) and Tidwell et al. (1980).

Collections of simuliids and ceratopogonids were also made from human volunteers to establish whether any natural filarial infection was present. These flies were immediately preserved in 80% ethanol following capture, and were later processed and examined in the laboratory using the method detailed under experimental infection.

RESULTS

Prevalence of mansonelliasis - In order to select a volunteer with a high microfilaraemia for the experimental infection work, individuals, who were known to be positive in the 1995 survey, were re-examined, even if they had been treated with ivermectin. Table II shows the results of this survey. Although our blood sampling was not done quantitatively the results clearly show that despite ivermectin treatment four individuals out of the 11 treated were infected with *M. ozzardi*.

Simuliid and ceratopogonid species - The following species of *Simulium* were recorded in the area: *S. exiguum* Roubaud, *S. dinellii* (Joan), *S. romanai* Wygodzinsky and *S. woffhuegeli* (Enderlein). Small numbers of *S. exiguum* and *S. woffhuegeli* were collected biting the ears and belly of horses, but only the occasional *S. dinellii* and moderate numbers of *S. exiguum* bit man. The Ceratopogonidae recorded from the area were: *C. paraensis* (Goeldi) and *Forcipomyia stylifera* Lutz

TABLE II
Infection of people with *Manzolla ozzardi* at Normenta, Jujuy Province, Argentina

Sex	Age	Resident in area (years)	Treated with ivermectin in Dec.95	Current test 1998
Tested in 1995 (infected)				
M	9	9	Yes	-
M	40	33	No	+
M	47	47	Yes	-
M	49	49	Yes	+
M	64	45	Yes	-
M	70	70	Yes	+
M	74	74	No	+
M	74+	74+	Yes	+
F	10	10	Yes	-
F	27	5	Yes	+
F	41	41	Yes	-
F	53	53	Yes	-
F	60	60	Yes	-
Tested in 1995 (not infected)				
M	51	6	No	+
Not tested in 1995				
M	7	7	-	-
M	20	20	-	+
M	34	34	-	-
M	42	25	-	-
M	50	5	-	+
M	56	56	-	+
M	62	62	-	+
M	70	70	-	+
M	79	50	-	+
M	80	80	-	+
F	13	13	-	-
F	17	9	-	-
F	19	0.5	-	-
F	42	8	-	+
F	42	9	-	-
F	57	57	-	-
F	67	67	-	+

biting horses in low numbers, *C. lahillei* (Iches) biting cattle in low numbers and *C. lahillei* (263) and *C. paraensis* (46) biting man.

Infection of flies - Only *C. lahillei* and *S. exiguum* were collected in large enough numbers to analyse the development of *M. ozzardi*. There was a considerable difference in mean ingestion rates of the two species that could not be explained by the small size of the former species relative to *S. exiguum*. *C. lahillei* only ingested 3.2 microfilariae [mean (log + 1)] per fly (range 0-17, n=14) compared to 49.6 [mean (log+1)] (range 15-186, n= 12) in *S. exiguum*. A single *S. dinellii* collected replete from the bait contained 42 microfilariae. Table III gives details of the experimental infection of *C. lahillei* with *M. ozzardi*. Of 134 flies experimentally infected 62.7%

(84) later showed 395 developing filariae. L3 larvae were first found on post infection day (PID) 3, probably resulting from a natural infection in these wild caught flies as is possibly the case on PID 5. The largest numbers of L3 larvae found were 14 in the thorax and eight in the head. The majority of parasites had reached the final stage in samples from PID 6-9 showing that it has a synchronous cycle. The mean (log+1) production of L3 larvae during PIDs 6-9 was 1.7, signifying that 53.1% of the 3.2 microfilariae ingested per fly developed to the infective stage. Fewer microfilariae ingested by *S. exiguum* developed in the fly (Table IV) despite a 15 times higher intake of parasites than *C. lahillei*. Only 26.9% (21) of the 78 flies infected developed filariae. The highest number of L3 larvae found in

TABLE III
Experimental infection of *Culicoides lahillei* with *Mansonella ozzardi* in Argentina

Post infection day	Flies dissected			Parasites (% of each stage)				
	Total	No. infected	% infected	Total no.	mff	L1	L2	L3
1	4	3	75	3	100	0	0	0
2	2	2	100	4	0	100	0	0
3	5	2	40	7	0	71.4	0	28.6
4	5	1	20	1	0	0	100	0
5	5	4	80	22	0	68.2	27.3	4.5
6	6	3	50	9	0	0	0	100
7	15	10	66.7	43	0	0	0	100
8	52	34	65.4	194	0	0	0.5	99.5
9	40	25	62.5	112	0	0	2.7	97.3
Tota	134	84	62.7	395				

Mean (log +1) microfilaria intake 3.2

TABLE IV
Experimental infection of *Simulium exiguum* with *Mansonella ozzardi* in Argentina

Post infection day	Flies dissected			Parasites (% of each stage)				
	Total	No. infected	% infected	Total no.	mff	L1	L2	L3
1	16	8	50	30	100	0	0	0
2	2	0	0	-	-	-	-	-
3	4	1	25	5	0	100	0	0
4	8	1	12.5	4	0	0	100	0
5	12	6	50	30	3.3	93.3	3.3	0
6	10	0	0	-	-	-	-	-
7	6	2	33.3	2	0	0	50	50
8	19	3	15.8	6	16.7	0	66.6	16.7
9	1	0	0	-	-	-	-	-
Total	78	21	26.9	77				

Mean (log + 1) microfilaria intake 49.6

an individual fly was one and this was in the head. Development occurred to the L3 stage by PID 7, but the presence of few L3 larvae compared to microfilariae and L2 larvae on PID 8 indicated that the cycle was not synchronous. Of the mean (log +1) microfilarial intake of 49.6 only a mean (log + 1) of 0.04 (0.8%) L3 larvae developed. Despite low biting densities five *C. paraensis* were experimentally infected and survived to PID 7 – two flies each contained an L3 larva of *M. ozzardi* in the head. Similarly, only one *S. dinellii* of eight experimentally infected contained filariae, five L2 stage larvae.

Natural infections of flies with filariae were as follows. Of 263 *C. lahillei* biting man two specimens each contained an L1 larva giving a natural infection rate with filariae of 0.7% and 1.03% (one fly with a single L1 larva) of 97 *S. exiguum* was infected.

DISCUSSION

During the selection of a volunteer for the experimental infection study four of the 11 individuals treated two years ago with ivermectin in December 1998 (Table II) were again positive for microfilariae of *M. ozzardi* in the peripheral blood. Little is known about the effects of ivermectin on this parasite (Campbell 1991), except that Nutman et al. (1987) in a comprehensive study on one infected individual observed that microfilariae had disappeared from the blood for a nine month period following a single treatment with ivermectin (140 µg/kg body weight). It is probable that the drug had suppressed production of microfilariae for this period but not killed the female worm, as occurs in onchocerciasis. Ivermectin is an efficient microfilaricide for *O. volvulus* but has to be administered at least once annually (150 µg/kg body weight) for many years because new microfilariae

produced by female worms will appear if only a single dose is given (WHO 1995).

It is not known exactly how many ceratopogonid and simuliid species occur in the area of Finca Normenta because no comprehensive seasonal survey has been made. Previous works in this region of Argentina (S Coscarón, unpub. data, Romaña & Wygodzinsky 1950) indicate that *S. dinellii* bites man in greater numbers in the wet season and that the anthropophilic species, *S. jujuyense* Paterson & Shannon and the zoophilic species *S. lahillei* (Paterson and Shannon) are also present. The data presented clearly show that *M. ozzardi* microfilariae will develop to maturity in *C. lahillei*, *C. paraensis* and *S. exiguum*. The presence of L2 larvae in *S. dinellii* on PID 5 may have resulted from the experimental infection with *M. ozzardi*, but could equally have been the result of the fly having had a previous blood meal on another animal that was infected with another filarial species; both *O. cervicalis* (Roux et al. 1984) and *O. gutturosa* (Mancebo & Roux 1986) have been recorded in horses and cattle respectively in Argentina. *S. dinellii* was discounted as a vector in Tucumán province to the north of Finca Normenta by Romaña and Wygodzinsky (1950) because of the failure of microfilariae to develop beyond this stage in flies that had fed on a volunteer infected with *M. ozzardi*. From our data *C. lahillei* is clearly a vector in the area because a high proportion of microfilariae developed to infective larvae in the experimental infection and on PID 3 the infective larvae present must have developed from an infection already carried by the fly when it received its experimental infective feed. *C. paraensis* is also probably a vector. Despite ingesting 15 times more microfilariae than *C. lahillei* development of the parasite in *S. exiguum* was poor, but did continue until the L3 stage. Consequently, *S. exiguum* probably sporadically transmits the parasite in this locality. Only L1 larvae were recorded from naturally infected flies and so it was not possible to identify them to species.

Only sufficient data are available for *C. lahillei* and *S. exiguum* for comparisons with other ceratopogonid and simuliid vectors of *M. ozzardi* in Latin America. Our data have shown that in Argentina *C. lahillei* is an equally efficient host to *M. ozzardi* as *C. furens* in Haiti (Lowrie & Raccurt 1981) and *C. phlebotomus* in Trinidad (Nathan 1981) in terms of infection rates, rapidity of the development cycle and high production of infective L3 larvae from ingested microfilariae. However, not all *Culicoides* species are efficient hosts to *M. ozzardi*. *C. insinuatus* was a less efficient host to this filaria in Colombia (Tidwell & Tidwell 1982), while Lowrie and Raccurt (1981) and

Raccurt and Lowrie (1981) showed *C. furens* to be a more effective host than *C. barbosai* and *Leptoconops bequaerti* to be the least effective in Haiti. *S. exiguum* in Finca Normenta was a relatively poor host to *M. ozzardi* because of low percentage development of microfilariae to infective larvae and an asynchronous development cycle. This was not the case in Amazonian simuliid species (*S. amazonicum*, *S. argentiscutum* and *S. oyapockense* s.l. [as *Simulium* sp., *S. minusculum*, *S. sanchezii*], reviewed in Yarzabal et al. (1985), which were all more effective hosts in terms of these parameters. It has been shown by Shelley et al. (1987) that *S. oyapockense* s.l. in the Brazilian Amazon is a relatively poor host to *O. volvulus* because of the damage caused by its cibarial teeth to microfilariae during ingestion, and that this was a major factor in determining vectorial competence in vector simuliid species in Latin America (Shelley 1988, 1994). Generally, species with unarmed cibaria are more effective hosts to *O. volvulus* than those with armed cibaria and the same must be true for *M. ozzardi*. All the species cited in Yarzabal et al. (1985) possess cibarial teeth, whereas *S. exiguum* at Finca Normenta does not. This would therefore suggest that this simuliid population is a poor host to *M. ozzardi* because of rapid formation of the peritrophic membrane or because of an inhibitor in the thoracic musculature as is seen in some simuliid species that transmit *O. volvulus* in Africa and Latin America (Shelley 1988, 1994). Host capacity variations also occur at the level of cytospecies. *S. exiguum* is a species complex in which individual cytospecies show different capacities to act as hosts to *O. volvulus* in Ecuador (Shelley et al. 1990) and in which some populations (Venezuela) are actually poor hosts (Shelley 1994).

Various data are now available that further refute the theory that two strains of *M. ozzardi* occur, the Caribbean with a ceratopogonid vector and the South American with a simuliid vector. Tidwell and Tidwell (1982) showed that *S. amazonicum*, *S. argentiscutum* and *C. insinuatus* occurred sympatrically in the Colombian Amazon and that all three acted as intermediate hosts to *M. ozzardi*, but the ceratopogonid species was less competent as a host than the two simuliid species. Similar findings on these three species were reported by Shelley et al. (1980) from the Brazilian Amazon, except that none of the few *C. insinuatus* experimentally infected with microfilariae of *M. ozzardi* produced developing parasites. The evidence that we present here from the Argentina focus shows that species from both families are able to host this parasite but that ceratopogonids are clearly the more effective hosts. Studies on the parasite itself show that popu-

lations from Haiti and the Colombian Amazon are almost morphologically identical (Kozek & Raccurt 1983) and that the DNA of the Argentine population from Finca Normenta is identical with that from the Brazilian Amazon (R Moraes Hojas, pers. comm.). On the existing evidence, it is therefore more probable that *M. ozzardi* is a widespread monomorphic species in Latin America that may be transmitted by ceratopogonid and/or simuliid species to varying degrees of competence depending on the vector population concerned.

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REFERENCES

- Biagi F, Tay J, Biagi AM 1958. Observaciones sobre mansonelosis en la Península de Yucatán. V. *Culicoides furens* como transmisor. *Medicina* 38: 377-379.
- Bruijning CFA 1957. Notes on the common species of *Culicoides* (Diptera: Ceratopogonidae) from Surinam in relation to *ozzardi*-filariasis. *Doc Med Geog Trop* 9: 169-172.
- Buckley JJC 1934. On the development, in *Culicoides furens* Poey, of *Filaria* (= *Mansonella*) *ozzardi* Manson, 1897. *J Helminthol* 12: 99-118.
- Campbell WC 1991. Ivermectin as an antiparasitic agent for use in humans. *Ann Rev Microbiol* 45: 445-474.
- Jörg ME 1983. Filariasis por *Mansonella ozzardi* (Manson 1897) Faust 1929 en la Argentina. *Prensa Méd Argent* 70: 181-192.
- Kozek WJ, Raccurt C 1983. Ultrastructure of *Mansonella ozzardi* microfilaria, with a comparison of the South American (simuliid transmitted) and the Caribbean (culicoid transmitted) forms. *Tropen Parasit* 34: 38-53.
- Lowrie RC, Raccurt C 1981. *Mansonella ozzardi* in Haiti. II. Arthropod vector studies. *Am J Trop Med Hyg* 30: 598-603.
- Lowrie RC, Raccurt C 1984. Assessment of *Culicoides barbosai* as a vector of *Mansonella ozzardi* in Haiti. *Am J Trop Med Hyg* 33: 1275-1277.
- Lowrie RC, Raccurt CP, Eberhard ML, Katz SP 1983. Assessment of *Leptoconops bequaerti* as a potential vector of *Mansonella ozzardi* in Haiti. *Am J Trop Med Hyg* 32: 1013-1015.
- Mancebo OA, Roux JP 1986. Primera determinación de *Onchocerca gutturosa* (Neumann, 1910) en la República Argentina: 1. Prevalencia de microfilarias en bovinos de la provincia de Formosa. Comunicación. *Vet Arg* 3: 165-166.
- Minoprio JL, Jörg ME, Marrelli VR 1988. *Mansonella ozzardi*, Faust 1922. Paciente infestado con madera procedente del norte argentino. *Prensa Méd Argent* 73: 330-332.
- Moraes MAP, Shelley AJ, Luna Dias APA 1985. *Mansonella ozzardi* no Território Federal de Roraima, Brasil. Distribuição e achado de um novo vetor na área do Rio Surumu. *Mem Inst Oswaldo Cruz* 80: 395-400.
- Nathan MB 1981. Transmission of the human filarial parasite *Mansonella ozzardi* by *Culicoides phlebotomus* (Williston)(Diptera: Ceratopogonidae) in coastal north Trinidad. *Bull Ent Res* 71: 97-105.
- Nathan MB, Tikasingh ES, Munroe P 1982. Filariasis in Amerindians of western Guyana with observations on transmission of *Mansonella ozzardi* by a *Simulium* species of the *amazonicum* group. *Tropenmed Parasit* 33: 219-222.
- Nelson GS, Pester FRN 1962. The identification of infective filarial larvae in Simuliidae. *Bull WHO* 27: 473-481.
- Nelson GS, Davies JB 1976. Observations on *M. ozzardi* in Trinidad. *Trans R Soc Trop Med Hyg* 70: 16-17.
- Nutman TB, Nash TE, Ottesen EA 1987. Ivermectin in the successful treatment of a patient with *Mansonella ozzardi* infection. *J Infect Dis* 156: 662-665.
- Petersen JL, Bawden MP, Wignall FS, Latorre CR, Johnson CM, Miranda C 1984. *Mansonella ozzardi* en el Darien (Panama). *Rev Méd Panama* 9: 236-246.
- Raccurt CP, Lowrie RC 1981. *Mansonella ozzardi* (Manson, 1897) et ses vecteurs en Haiti (Grandes Antilles). *Lyon Médical* 245: 135-139.
- Remondegui C, Safarov G, Ripoll CM, Arce de Hamity MG, Neder de Roman LE, Esquivel O 1988. *Mansonella ozzardi*: estudio clínico epidemiológico de un foco endémico en la provincia de Jujuy. *Acta Infectología* 4: 3-13.
- Romaña C, Wygodzinsky P 1950. Acerca de la transmisión de *Mansonella ozzardi* (Manson) (*Filaria tucumana* Biglieri y Araoz). *Anales del Instituto de Medicina Regional* 3: 29-34.
- Roux JP, Mancebo OA, Bulman GM 1984. Primera determinación de *Onchocerca cervicalis* (Railliet y Henry, 1910) en la República Argentina: 1. Prevalencia de microfilarias en equidos del nordeste subtropical. *Therios* 3: 372-378.
- Shelley AJ 1988. Vector aspects of the epidemiology of onchocerciasis in Latin America. *Ann Rev Ent* 33: 337-366.
- Shelley AJ 1994. Factors affecting filarial transmission by simuliids. *Adv Disease Vector Research* 10: 183-214.
- Shelley AJ, Charalambous M, Arzube M 1990. *O. volvulus* development in four *S. exiguum* cytospecies in Ecuador. *Bull Soc Française Parasitol* 8: 1145.
- Shelley AJ, Luna Dias APA, Moraes MAP 1980. *Simulium* species of the *amazonicum*-group as vectors of *Mansonella ozzardi* in the Brazilian Amazon. *Trans R Soc Trop Med Hyg* 74: 784-788.
- Shelley AJ, Luna Dias APA, Moraes MAP, Procnunier WS 1987. The status of *Simulium oyapockense* and *S. limbatum* as vectors of human onchocerciasis in Brazilian Amazonia. *Med Vet Entom* 1: 219-234.

- Shelley AJ, Shelley A 1976. Further evidence for the transmission of *Mansonella ozzardi* by *Simulium amazonicum* in Brazil. *Ann Trop Med Parasit* 70: 213-217.
- Taranto NJ, Castelli E 1988. Detección de un foco de microfilariasis en el noroeste argentino. *Rev Arg Microb* 20: 49-51.
- Tidwell MA, Tidwell MA 1982. Development of *Mansonella ozzardi* in *Simulium amazonicum*, *S. argentiscutum*, and *Culicoides insinuatus* from Amazonas, Colombia. *Am J Trop Med Hyg* 31: 1137-1141.
- Tidwell MA, Tidwell MA, Muñoz de Hoyos P 1980. Development of *Mansonella ozzardi* in a black fly species of the *Simulium sanguineum* group from eastern Vaupés, Colombia. *Am J Trop Med Hyg* 29: 1209-1214.
- Undiano C 1971. Importancia y actualización del nuevo concepto de la patogenicidad de la mansoneliasis. *Segundas Jornadas Argentinas de Entomoepidemiológicas Argentinas* 2: 467.
- WHO 1995. Onchocerciasis and its control. *WHO Tech Rep Ser* 852: 1-103.
- Yarzabal L, Basáñez MG, Ramírez-Pérez J, Ramírez A, Botto C, Yarzabal A 1985. Experimental and natural infection of *Simulium sanchezi* by *Mansonella ozzardi* in the Middle Orinoco region of Venezuela. *Trans R Soc Trop Med Hyg* 79: 29-33.